



http://jkmu. kmu. ac. ir/

The Radioprotective Effect of Magnesium Sulfate and Vitamin A on Radiation-induced Micronuclei and the Expression of NOX4 in Bone Marrow Cells of Mice

Mohammad Mirdoraghi¹, Vahid Changizi², Seyed Abolghasem Haeri³, Zahra Rajabi⁴, Peyman Amini², Sakineh Abbasi^{5*}

1. Department of Medical Physics and Biomedical Engineering, Tehran University of Medical Sciences, Tehran, Iran

- 2. Department of Radiology and Radiotherapy Technology, School of Allied Health Sciences, Tehran University of Medical Sciences, Tehran, Iran
- 3. Nuclear Science and Technology Research Institute, Tehran, Iran
- 4. Zoonosis Research Center, Tehran University of Medical Sciences, Tehran, Iran

5. Department of Medical Laboratory Sciences, Faculty of Allied Medicine, Tehran University of Medical Sciences, Iran



ABSTRACT

Background: Radioprotectors are used to neutralize the effects of free radicals caused by ionization radiation. In this study, the radioprotective effects of magnesium sulfate and vitamin A on bone marrow cells of mice were evaluated by micronucleus assay and changes in the expression of NOX4 gene.

Methods: The mice were randomly divided into 12 groups. The mixture of drugs was injected into mice by intraperitoneal injection 2 hours before the irradiation. The dose rate was 50 cGy/min at SSD (source to surface distance) 100 cm and field size of 10×10 cm². Twenty four hours after 2 Gy irradiation by LINAC, the mice were sacrificed by cervical dislocation. Then, several microscopic slides were prepared for each sample to evaluate the number of micronucleus in polychromatic erythrocytes (PCEs). In addition, the expression of NOX4 was evaluated by Real-time PCR. Data were analyzed through SPSS 19 and the mean of groups was compared to each other using one-way ANOVA.

Results: There was a significant difference between mean mnPCEs in the treatment (drugs + radiation) groups compared to the 2 Gy group (P=0.01). The expression level of NOX4 gene was significantly lower in groups receiving the combinations of vitamin A and magnesium sulfate compared to the 2 Gy group (P =0.01). The calculated dose reduction factor (DRF) demonstrated DRF=2.58 for 2Gy.

Conclusion: The results of this study indicated that the combination of vitamin A and magnesium sulfate, possibly with an antioxidant mechanism, removes the deleterious effects of free radicals caused by ionizing radiation on bone marrow cells.

Keywords: Micronucleus assay, Magnesium sulfate, NOX4, Radiation-protective agents, Vitamin A

Received: 03. 08. 2021

*Correspondence: Sakineh Abbasi; Email: sakineh4612004@yahoo.com

Published by Kerman University of Medical Sciences

Citation: Mirdoraghi M, Changizi V, Haeri SA, Rajabi Z, Amini P, Abbasi S. The radioprotective effect of magnesium sulfate and vitamin A on radiationinduced micronuclei and the expression of NOX4 in bone marrow cells of mice. Journal of Kerman University of Medical Sciences 2022; 29(3): 237-245. doi: 10.22062/JKMU.2022.91947

Accepted: 12. 11. 2021

Introduction

I onizing radiation (IR) is one of the most threatening natural, occupational and medical agents that can bring about serious health damages (1). Although the dose of the occupational radiation is generally low, it could increase exceedingly due to the radiological accidents at nuclear reactors (2-4), exposure to radioactive waste (5), outcomes of nuclear bombing (6), the industrial accidents in the course of mining and processing of radioactive substances (7). In these cases, the risk of exposure to the great amount of radiation can reach a significant extent not only for radiation employees but also for personnel engaging in emergency response.

Clinical application of IR is broadly used for the treatment of a wide variety of cancers as a part of therapy. Half of all cancer patients are expected to undergo radiotherapy at some point in in the process of their cancer treatment. Previous studies have shown that augmenting the cumulative radiation dose by 10-20% can completely eradicate some tumors (8). However, high cumulative radiation dose can harm healthy tissues surrounding the tumor and therefore brings about side effects.

In order to protect the normal tissues from detrimental effects of IR, many radioprotectors and mitigators have been developed (9-12). Mitigators are used to decrease toxicity and applied even after irradiation (13,14). Radioprotectors are substances that are able to diminish the harmful effects of radiation on normal tissues and they should be present in the tissues before or at the time of irradiation (15).

Radioprotective agents that have been so far suggested are categorized into three groups; thiol compounds that are capable to neutralize the generated free radicals by radiation, cytokines and growth factors that alter cellular reaction to radiation by moderating communication between immune cells and other kinds of cells. and herbal extracts and natural antioxidant (16). The most common and effective radioprotectors are thiol substances. Amifostine, a thiol compound, is the only radioprotector that has been approved by the US food and drug administration (FDA). Nonetheless, it has adverse effects including vomiting, nausea and hypotension. Due to the toxicity of amifostine, its applications in clinical trials is considerably low compared to that in animal investigations (17). Another group of radioprotectors that have recently been considered is natural and herbal

antioxidants, such as melatonin, vitamin C, flavonoids and so on. These natural compounds have fewer radioprotective effects and lower adverse effects compared to the thiol compounds. One approach to increase the effectiveness of radioprotectors and diminish their toxicity is to investigate natural radioprotectors, such as vitamins and magnesium sulfate (18). Vitamin A is one of the fat-soluble vitamins and is essential for the growth of the body, the proper functioning of the immune system, and the prevention of infections (19). Vitamin A does not exist purely in plant sources, but in its precursors, carotenes, in various forms. The most common precursor of vitamin A is beta-carotene, some of which is converted to vitamin A in the body. Betacarotene effectively reduces the production of trichloromethyl peroxyl radicals and protects the membrane against lipid peroxidation (20). Previous studies have found that magnesium has antioxidant effects, and its ability to reduce and scavenge free radicals has been of interest to researchers (21-23). Magnesium inhibits nicotinamide adenine dinucleotide phosphate oxidase (NOX), which increases the production of oxidized free radicals. As a result, magnesium can directly inhibit the production of free radicals or scavenge free radicals (24, 25).

There are several assays to assess the efficacy of radioprotectors, such as Micronucleus (MN) assay, dicentric assay and gene expression method (26). MN is formed by the break of chromosomes at the anaphase stage of mitosis in immature polychromatic erythrocytes (PCE). The development of immature erythrocytes or PCE into mature erythrocytes or normochromatic erythrocytes (NCE) takes about 6-7 cell divisions. Using the MN method, chromosomal damages that change the number and shape of chromosomes can be counted (27).

Although the dicentric assay is currently the gold standard for biodosimetry, it is not possible to use this technique at high mass casualty rates without automatic dicentric methods. Even with automated counts, dicentric counts require cell division, so it takes at least 3 days for the results to be ready (28). Since gene expression does not require cell division, it can estimate the dose within hours (29). Exposure of human cells to environmental stress including IR activates several cellular signal pathways and rapidly leads to complex patterns of gene expression. Expression does and the kind of oxidative stress.

Gene expression changes may continue for several days after radiation exposure. These gene expression changes can be used to estimate radiation dose and radiation damages (30). In addition, changes in the expression of specific genes including NOX4 in the control and case groups can approximately estimate the radioprotective property of drugs.

NOX gene encodes a member of the NOX enzyme family that acts as an oxidase. NADPH is a catalytic subset of the oxidase complex of nicotinamide adenine dinucleotide phosphate oxidase, an NADPH membrane enzyme complex that faces the extracellular space. This encoded protein binds to non-phagocytic cells, where it acts as an oxygen sensor and catalyzes molecular oxygen to reactive oxygen species (ROS). The ROS produced by this protein has been implicated in various biological functions including signal transduction, cell differentiation and tumor cell growth (31).

The role of NOX4 in many cancers, such as glioma, melanoma, and thyroid cancer is also significant. It has been shown that NOX4 contributes to the progression of metastasis in various cancers (32). Other studies have shown that NOX4 regulates the cell cycle, decreases proliferation, and increases cell apoptosis (33, 34). Measuring changes in the expression of this gene in the groups of animals given radioprotective drugs compared to the control group, along with the MN method can give us more information to more accurately estimate the radioprotective properties of the substances.

Given the adverse effects of radiation in radiotherapy and the lack of appropriate radioprotectors, such as amifostine due to its side effects, further research is needed to achieve radioprotective compounds with fewer side effects and higher radioprotective properties. Therefore, the aim of this study was to investigate the radioprotective effect of magnesium sulfate and vitamin A in combined doses to find out whether magnesium sulfate and vitamin A have synergistic effects on the radioprotective activity of each other.

Materials and Methods Grouping of animals

In this study, 6-7 weeks old NMRI male mice were used. They were kept at a suitable temperature and 12/12 light cycle. The mice were categorized into 12 groups so that each group contained 5 mice (Table 1). In addition, the ethical code from the Ethical Committee of Tehran University of Medical Sciences was obtained, animals were also treated in accordance with Guide for the Care and Use of Laboratory Animals (8th edition, National Academies Press) (35). The Ethical Committee for medical Research at Tehran University of Medical Science, endorsed this research [ethical code IR.TUMS.SPH.REC.1396.4098].

Prescription of drugs

The radioprotective drugs were Vitamin A (DarouPakhsh Pharmaceutical Co, Tehran, Iran) and magnesium sulfate (Merck, Germany, pa). The mice were given 9 different combined doses of vitamin A (100, 200 and 400 mg / kg) and magnesium sulfate (75, 150 and 300 mg/kg) to determine the optimal dose. Vitamin A was dissolved individually in ethanol (%5). Then different combined doses of drugs were injected intraperitoneally into the mice by insulin syringes. Also, ethanol (%5) that is the solvent of vitamin A was injected into a group of mice to realize the difference between control group (group A) and ethanol group (group B). Mice were placed in standard irradiation cages and exposed to 2 Gy (whole-body irradiation) of X radiation by 10 MV x-ray beams from a linear accelerator (Varian 2100 CD). The dose rate was 50 cGy/min at SSD (source to surface distance) = 100 cm and field size of $10 \times 10 cm^2$ (36).

Bone marrow sampling

Twenty four hours after irradiation, the animals were anesthetized and sacrificed by cervical dislocation. Then, both femoral bones were removed, the bone marrow of each femur was extracted by 1 cc fetal bovine serum (FCS) from the lower end of the femur and transferred into the microtube. For each mouse, the bone marrows of both femurs were extracted into two microtubes. One microtubule for MN test and the other for evaluation of gene expression.

MN test

The bone marrow cells were centrifuged at 2000 rpm for 6 min at 4 $^{\circ}$ C; then a series of the bone marrow cells was placed at -70 $^{\circ}$ C for RNA extraction and the other for the MN technique. The cells were afterward transferred to the microscopic slides and fixed for 5 minutes using the methanol solution, dried by exposure to open air for 24 hours and stained with May-Grünwald-Giemsa staining solution. To determine the number of MN in each sample, 1,000 PCEs were

counted by a Y100 Nikon microscope with $100 \times$ objective lens (37,38).

Gene expression method

Generally, gene expression consists of three stages; extraction of RNA from bone marrow cells, cDNA synthesis (RT-PCR) and Real-time PCR. To evaluate the expression of NOX genes in tumor tissues, RNA was extracted using RNeasy Mini Kit (Qiagen). Also, its quality and quantity were assessed using agarose gel (1.5%)and spectrophotometer (Thermo ScientificTM NanoDrop-1000), respectively. To perform cDNA synthesis (RT-PCR), 2 micrograms RNA were converted into cDNA by SuperScript II reverse transcriptase (Invitrogen); in addition, Oligo (dT)₁₅ primer (Roche) that adhere to the poly A of mRNAs was used. To carry out Realseveral materials time PCR, including characteristic primers of Nox4 [Table 2], Light Cycler[®] FastStart DNA Master^{PLUS} SYBR Green I, a Light Cycler Real-time machine (Roche) were used. GAPDH gene was used as an internal standard (Table 2). To compare the relative quantities (RQ) of gene expression between treatment groups and control group, the fold change of the gene was calculated using the comparative CT method, known as the $2^{-\Delta\Delta Ct}$ method (39).

Statistical analysis

Normal distribution of data was performed using histogram in SPSS 16; the mean MN and gene expression were expressed as Mean \pm SE. Then, to compare means of MNPCEs and means of gene expression changes in groups, one way ANOVA was used. Also, the differences between the means of different groups were determined by Tukey's Post Hoc Test (P <0.05).

Results

Statistical analysis showed that the combination of vitamin A and magnesium sulfate had a significant radioprotective effect. The MnPCEs/1000PCEs was 98.66 ± 3.05 in 2 Gy x-ray group; however, it was 34 ± 2 in group H (table 3). There was a significant difference (P=0.01) between mean mnPCEs in the treatment (drugs + radiation) groups compared to the 2 Gy group C (table 3). The calculated dose reduction factor (DRF) demonstrated DRF=2.58 for 2Gy (table 3).

Mean \pm standard error of mean (SEM) of NOX4 gene expression was 15.94 ± 0.011 in 2 Gy x-ray group; although, it decreased to

 0.76 ± 0.052 in group H (table 4). The expression level of NOX4 gene was significantly lower in groups receiving the combinations of vitamin A and magnesium sulfate compared to the 2 Gy group C (P =0.01). But, the differences in the mean expression of Nox4 in the group D, group E, and group H compared to the group C were greater (P= 0.001). The highest difference in Nox4 gene expression was observed between 2Gy+200 mg/kg vit A+150 mg/kg mgso4 group and 2 Gy Group C (table 4).

Table 1. The division of animals into the 12 studied groups

Group code	Dose of X	Dose of drugs
	radiation (Gy)	(mg/kg)
Α	0	0
В	0	5 cc ethanol
		(5%)
С	2	0
D	2	100 vit A + 75
		$mgso_4$
Ε	2	100 vit A + 150
		$mgso_4$
F	2	100 vit A + 300
		$mgso_4$
G	2	200 vit A + 75
		$mgso_4$
Н	2	200 vit A + 150
		$mgso_4$
I	2	200 vit A + 300
		$mgso_4$
J	2	400 vit A + 75
		$mgso_4$
K	2	400 vit A + 150
		$mgso_4$
L	2	400 vit A + 300
		$mgso_4$

Table 2. Forward and reverse primers of Nox4 and		
GAPDH gen	es	
Primer	Sequence	
GAPDH F	5-CCCTTAAGAGGGATGCTGCC-3	
GAPDH R	5-TACGGCCAAATCCGTTCACA-3	
NOX4 F	5- TTGCCTGGAAGAACCCAAGT -3	
NOX4 R	5- TCCGCACAATAAAGGCACAA -3	

Table	3.	Mea	n±SE	freque	ncie	s of
MnPCEs	s/1000F	CEs in	n bone	marrow	in	various
groups, 2	24 hour	s after 2	2 Gy of	X radiation	on	

Groups	MnPCEs/1000PCEs	Dose Reduction Factor (DRF)
A	20 ± 2.16	
В	31.80 ±3 .27	
С	98.66 ± 3.05	
D	53.66 ± 4	1.83
Е	33 ± 2	2.98
F	33 ± 2	2.98
G	37.33 ± 3	2.64
Н	34 ± 2	2.90
I	34 ± 4	2.90
J	45 ± 4	2.19
K	49.33 ± 8	2
L	35 ± 1	2.81

Table 4. Mean \pm standard error of mean (SEM) of
NOX4 gene expression in different groups

Groups	Fold change
С	15.94±0.011
D	1.53±0.011
Ε	3.02±0.25
F	1.59±0.20
G	4.27±0.018
Н	0.76±0.052
I	2.85±0.023
J	2.85±0.023
К	3.52±0.33
L	8.81±1.78

Discussion

In the present study, the expression of NOX4 gene was highest in the group receiving only 2 Gy X-ray, indicating that irradiation increases the expression of NOX4 gene. In Collins-Underwood *et al.* study, the expression of NOX gene increased in the brain endothelial cells of rats after 10 Gy irradiation (40). Moreover, Pazhanisamy et al. also found that inhibition of Nox gene by the diphenylene iodine after wholebody irradiation of 6.5 Gy reduces the genomic instability of the hematopoietic system (41). Irradiation has been shown to induce chronic oxidative stress response to NOX activity in rat cells. This finding suggests that reduction in the expression of NOX could be a sign of decline in radiation-induced DNA damage and cell death (42). Therefore, the result of our research is in line with the results of previous studies.

Some investigations have indicated that radioprotectors can decrease the expression of Nox4 gene. In Najafi *et al.* study, whole-body irradiation increased the expression of NOX4 gene and melatonin reduced the expression of NOX4 gene in target and non-target lung tissues (43). Moreover, Yang et al. figured out that the expression of Nox4 gene increased in radiotherapy of fibrosis, and treatment with magnesium isoglycyrrhizinate diminished the expression of Nox4 gene and protected healthy tissue against radiation (44). In addition, Jiang et al. found that X-ray irradiation of mice enhanced the expression of NOX4 gene and naringenin treatment reduced the expression of this gene and protected the animal's healthy tissues against radiation (45). Based on the results of the mentioned studies, it can be concluded that the reduction of NOX4 gene expression can be considered as a criterion to compare the protective effect of radioprotective agents.

The results of our study showed that injection of vitamin A and magnesium sulfate in the mixed doses can have a radioprotective effect against 2 Gy X-ray. In both methods, gene expression and MN assay, the differences between treatment groups and the 2 Gy X-ray group were significant. Based on the results of gene expression technique, the most effective combination of vitamin A and magnesium sulfate was 200 mg/kg and 150 mg/kg, respectively. Also, based on the results of MN technique, the most effective dose has a dose reduction factor (DRF) of 2.9, which is consistent with the results of the gene expression technique.

In general, the radioprotective mechanism of vitamin A is attributed to its antioxidant properties. Vitamin A plays a protective role against radiation by blocking the pathways of chain reactions initiated by free radicals (46). The most common precursor of vitamin A is beta-carotene, which effectively reduces the produced trichloromethyl peroxyl free radicals and protects the membrane against lipid peroxidation (47). Retinol scavenges free radicals by inhibiting peroxidation in a homogeneous methyl linoleate solution. To stabilize and neutralize peroxyl free radicals, vitamin A is oxidized by these free radicals, producing 5, 6-epoxy retinoic and ultimately stabilizing free radicals (48). Soybean oilsoluble vitamin A has been shown to protect healthy tissues from the radiation damage caused by internal radionuclides (49).

The radioprotective effect of magnesium sulfate in both MN assay and gene expression method was significant. Studies have shown that the insufficiency of magnesium increases the rate of oxidative cell death, and magnesium can augment the stability of DNA, the maintenance of enzymes involved in protein biosynthesis, gene transcription, protein production, and cell growth. Magnesium also plays a fundamental role in the structure and physiology of cells (50, 51). Magnesium has been shown to have a strong anti-inflammatory capacity as well as an antioxidant role against free radicals (52). Geiger et al. showed that magnesium plays a role in preventing oxidative stress so that insufficient magnesium increases blood pressure, glucose resistance, and insulin resistance (53). Since magnesium is a natural antagonist of calcium and also has antioxidant effects, it is likely that magnesium sulfate scavenges free radicals using antioxidant properties. Nonetheless, further research is needed to determine the extent of its radioprotective effect.

The greatest difference of means was observed between group H (2GY radiation + 200mg/kg Vit A + 150mg/kg mgso₄ and group C (2GY radiation+ no treatment). The results of this study show that the radioprotective effect of the combination of two drugs, vitamin A and magnesium sulfate, is promising. The optimal combination dose of the two drugs in wholebody irradiation is found in group H.

References

- 1. Mirdoraghi M, Einor D, Baghal-Asghari F, Esrafili A, Heidari N, Mohammadi AA, et al. Assess the annual effective dose and contribute to risk of lung cancer caused by internal radon 222 in 22 regions of Tehran, Iran using geographic information system. Journal of Environmental Health Science and Engineering. 2020; 18(1):211-20. doi: 10.1007/s40201-020-00454-3.
- Barquinero JF, Fattibene P, Chumak V, Ohba T, Della Monaca S, Nuccetelli C. et al. Lessons from past radiation accidents: Critical review of methods addressed to individual dose assessment of potentially exposed people and integration with medical assessment. Environment International. 2021; 146:106175. doi: 10.1016/j.envint.2020.106175.
- 3. Lee Y, Choi YY, Yang M, Jin YW, Seong KM. Risk perception of radiation emergency medical staff on low-dose radiation exposure: Knowledge is a critical factor. J Environ

There were some limitations in our work such as using only two radiobiological dosimetry methods to evaluate the radioprotective effects of the drugs due to low budget of the project. We recommend other researchers to carry out more experiments on the radioprotective effects of vitamin A and mgso₄ on tumoral and normal tissues using different doses of x-ray.

Conclusion

In summary, this study showed that the radioprotective effect of the combination of two drugs, vitamin A and magnesium sulfate, were relatively high for protection against 2 Gy X-ray. In addition, the expression of NOX4 gene and the number of mnPCEs in bone marrow cells increased by 2 Gy X-ray irradiation. The results of this study also suggest that the combination of magnesium and vitamin A on the expression of NOX4 gene may protect the bone marrow cells of mice against IR damage. Thus, a mixture of both vitamin A and magnesium sulfate can be used as a radioprotective agent in patients undergoing radiotherapy, occupational exposure, nuclear accidents and space travel.

Conflict of interest

Not declared.

Radioact. 2021; 227:106502. doi: 10.1016/j.jenvrad.2020.106502.

- Wang Z, Chen Z, Chen C, Ge D, Perrault D, Zucchetti M, et al. Quantitative safety goals for fusion power plants: Rationales and suggestions. International Journal of Energy Research. 2021; 45(6):9694-703. doi: 10.1002/er.6399.
- Wisnubroto DS, Zamroni H, Sumarbagiono R, Nurliati G. Challenges of implementing the policy and strategy for management of radioactive waste and nuclear spent fuel in Indonesia. Nuclear Engineering and Technology. 2021; 53(2):549-61. doi: 10.1016/j.net.2020.07.005.
- Bolton MB, Minor E. Addressing the ongoing humanitarian and environmental consequences of nuclear weapons: an introductory review. Global Policy. 2021; 12(1):81-99. doi: 10.1111/1758-5899.12892.

- Gorlenko NV, Leonova MS, Murzin MA. Occupational risks in the extraction and processing of mineral raw materials. IOP Publishing. 2020; 459(3):032023. doi: 10.1088/1755-1315/459/3/032023.
- 8. Linkous AG, Yazlovitskaya EM. Novel radiosensitizing anticancer therapeutics. Anticancer Res. 2012; 32(7):2487-99. PMID: 22753705.
- Agbele AT, Fasoro OJ, Fabamise OM, Oluyide OO, Idolor OR, Bamise EA. Protection against ionizing radiation-induced normal tissue damage by resveratrol: A systematic review. Eurasian J Med. 2020; 52(3):298-303. doi: 10.5152/eurasianjmed.2020.20143.
- Changizi V, Haeri SA, Abbasi S, Rajabi Z, Mirdoraghi M. Radioprotective effects of vitamin A against gamma radiation in mouse bone marrow cells. MethodsX. 2019; 6:714-7. doi: 10.1016/j.mex.2019.03.020
- Farhood B, Goradel NH, Mortezaee K, Khanlarkhani N, Salehi E, Nashtaei MS, et al. Melatonin as an adjuvant in radiotherapy for radioprotection and radiosensitization. Clin Transl Oncol. 2019; 21(3):268-79. doi: 10.1007/s12094-018-1934-0.
- Musa AE, Omyan G, Esmaely F, Shabeeb D. Radioprotective effect of hesperidin: A systematic review. Medicina (Kaunas). 2019; 55(7):370. doi: 10.3390/medicina55070370.
- Bijman R, Rossi L, Sharfo AW, Heemsbergen W, Incrocci L, Breedveld S, et al. Automated radiotherapy planning for patient-specific exploration of the trade-off between tumor dose coverage and predicted radiation-induced toxicity-a proof of principle study for prostate cancer. Front Oncol. 2020; 10:943. doi: 10.3389/fonc.2020.00943.
- Fitz Gerald TJ, Bishop Jodoin M, Laurie F, Riberdy C, Aronowitz JN, Bannon E, et al. Radiation therapy. Cancer: Prevention, Early Detection, Treatment and Recovery; 2019. P. 445-61.
- Mun G-I, Kim S, Choi E, Kim CS, Lee Y-S. Correction to: Pharmacology of natural radioprotectors. Arch Pharm Res. 2020; 43(2):272-4. doi: 10.1007/s12272-019-01194-1.
- Smith TA, Kirkpatrick DR, Smith S, Smith TK, Pearson T, Kailasam A, et al. Radioprotective agents to prevent cellular damage due to ionizing radiation. J Transl Med. 2017; 15(1):232. doi: 10.1186/s12967-017-1338-x.

- Kamran MZ, Ranjan A, Kaur N, Sur S, Tandon V. Radioprotective agents: strategies and translational advances. Med Res Rev. 2016; 36(3):461-93. doi: 10.1002/med.21386.
- Yahyapour R, Shabeeb D, Cheki M, Musa AE, Farhood B, Rezaeyan A, et al. Radiation protection and mitigation by natural antioxidants and flavonoids: Implications to radiotherapy and radiation disasters. Curr Mol Pharmacol. 2018; 11(4):285-304. doi: 10.2174/1874467211666180619125653.
- Huang Z, Liu Y, Qi G, Brand D, Zheng SG. Role of vitamin A in the immune system. J Clin Med. 2018; 7(9):258. doi: 10.3390/jcm7090258.
- 20. Fuchs-Tarlovsky V. Role of antioxidants in cancer therapy. Nutrition. 2013; 29(1):15-21. doi: 10.1016/j.nut.2012.02.014.
- Ciscomani-Larios JP, Sánchez-Chávez E, Jacobo-Cuellar JL, Sáenz-Hidalgo HK, Orduño-Cruz N, Cruz-Alvarez O, et al. Biofortification efficiency with magnesium salts on the increase of bioactive compounds and antioxidant capacity in snap beans. Ciência Rural. 2021; 51(6). doi: 10.1590/0103-8478cr20200442.
- 22. Fernández M, Marín R, Proverbio F, Ruette F. Effect of magnesium sulfate in oxidized lipid bilayers properties by using molecular dynamics. Biochem Biophys Rep. 2021; 26:100998. doi: 10.1016/j.bbrep.2021.100998.
- Mohammadi HR, Shamshirian A, Eslami S, Shamshirian D, Ebrahimzadeh MA. Magnesium sulfate attenuates lethality and oxidative damage induced by different models of hypoxia in mice. Biomed Res Int. 2020; 2020: 2624734. doi: 10.1155/2020/2624734.
- 24. Durlach J (2007) Overview of magnesium research: History and current trends. In Nishizawa Y, Mori H, Durlach J, (Eds.), New perspectives in magnesium research: Nutrition and health. London, Springer-Verlag. P. 3-10.
- Matović V, Buha A, Bulat Z, Đukić-Ćosić D, Miljković M, Ivanišević J, et al. Routedependent effects of cadmium/cadmium and magnesium acute treatment on parameters of oxidative stress in rat liver. Food Chem Toxicol. 2012; 50(3-4):552-7. doi: 10.1016/j.fct.2011.12.035.
- 26. Swartz HM, Williams BB, Flood AB. Overview of the principles and practice of biodosimetry. Radiat Environ Biophys. 2014; 53(2):221-32. doi: 10.1007/s00411-014-0522-0.

- Waldron D. Chromothripsis and micronucleus formation. Nature Reviews Genetics. 2015; 16(7):376-7. doi: 10.1038/nrg3970.
- Endesfelder D, Kulka U, Einbeck J, Oestreicher U. Improving the accuracy of dose estimates from automatically scored dicentric chromosomes by accounting for chromosome number. Int J Radiat Biol. 2020; 96(12):1571-84. doi: 10.1080/09553002.2020.1829152.
- 29. Macaeva E, Mysara M, De Vos WH, Baatout S, Quintens R. Gene expression-based biodosimetry for radiological incidents: assessment of dose and time after radiation exposure. Int J Radiat Biol. 2019; 95(1):64-75. doi: 10.1080/09553002.2018.1511926.
- 30. Knops K, Boldt S, Wolkenhauer O, Kriehuber R. Gene expression in low-and high-doseirradiated human peripheral blood lymphocytes: possible applications for biodosimetry. Radiat Res. 2012; 178(4):304-12. doi: 10.1667/rr2913.1.
- Gandara ACP, Torres A, Bahia AC, Oliveira PL, Schama R. Evolutionary origin and function of NOX4-art, an arthropod specific NADPH oxidase. BMC Evol Biol. 2017; 17(1):92. doi: 10.1186/s12862-017-0940-0.
- Jafari N, Kim H, Park R, Li L, Jang M, Morris AJ, et al. CRISPR-Cas9 mediated NOX4 knockout inhibits cell proliferation and invasion in HeLa cells. PLoS One. 2017; 12(1):e0170327. doi: 10.1371/journal.pone.0170327.
- Li J-M, Fan LM, George VT, Brooks G. Nox2 regulates endothelial cell cycle arrest and apoptosis via p21cip1 and p53. Free Radic Biol Med. 2007; 43(6):976-86. doi: 10.1016/j.freeradbiomed.2007.06.001.
- Tang C-T, Lin X-L, Wu S, Liang Q, Yang L, Gao YJ, et al. NOX4-driven ROS formation regulates proliferation and apoptosis of gastric cancer cells through the GL11 pathway. Cell Signal. 2018; 46:52-63. doi: 10.1016/j.cellsig.2018.02.007.
- Council NR. Guide for the care and use of laboratory animals. 8th. Washington, DC.: National Academies Press. 2011.
- Deloch L, Derer A, Hartmann J, Frey B, Fietkau R, Gaipl US. Modern radiotherapy concepts and the impact of radiation on immune activation. Front Oncol. 2016; 6:141. doi: 10.3389/fonc.2016.00141.
- 37. Kasamoto S, Masumori S, Hayashi M. In vivo micronucleus assay in mouse bone marrow and

peripheral blood. Methods Mol Biol. 2013; 1044:179-89. doi: 10.1007/978-1-62703-529-3_9.

- Van Miert E, Vanscheeuwijck P, Meurrens K, Gomm W, Terpstra PM. Evaluation of the micronucleus assay in bone marrow and peripheral blood of rats for the determination of cigarette mainstream-smoke activity. Mutat Res. 2008; 652(2):131-8. doi: 10.1016/j.mrgentox.2008.01.006.
- 39. Xia M, Sherlock J, Hegerich P, You X, Lee K, Walworth C, et al. Dataassist[™]-data analysis software for taqman[®] real-time PCR data. IMECS. 2010; 1:210-2.
- Collins-Underwood JR, Zhao W, Sharpe JG, Robbins ME. NADPH oxidase mediates radiation-induced oxidative stress in rat brain microvascular endothelial cells. Free Radic Biol Med. 2008; 45(6):929-38. doi: 10.1016/j.freeradbiomed.2008.06.024.
- Pazhanisamy SK, Li H, Wang Y, Batinic-Haberle I, Zhou D. NADPH oxidase inhibition attenuates total body irradiation-induced haematopoietic genomic instability. Mutagenesis. 2011; 26(3):431-5. doi: 10.1093/mutage/ger001.
- Wang Y, Liu L, Pazhanisamy SK, Li H, Meng A, Zhou D. Total body irradiation causes residual bone marrow injury by induction of persistent oxidative stress in murine hematopoietic stem cells. Free Radic Biol Med. 2010; 48(2):348-56. doi: 10.1016/j.freeradbiomed.2009.11.005.
- Najafi M, Shirazi A, Motevaseli E, Geraily G, Amini P, Tooli LF, et al. Melatonin modulates regulation of NOX2 and NOX4 following irradiation in the lung. Curr Clin Pharmacol. 2019; 14(3):224-31. doi: 10.2174/1574884714666190502151733.
- 44. Yang Q, Zhang P, Liu T, Zhang X, Pan X, Cen Y, et al. Magnesium isoglycyrrhizinate ameliorates radiation-induced pulmonary fibrosis by inhibiting fibroblast differentiation via the p38MAPK/Akt/Nox4 pathway. Biomed Pharmacother. 2019; 115:108955. doi: 10.1016/j.biopha.2019.108955.
- Jiang Y, You F, Zhu J, Zheng C, Yan R, Zeng J. Cryptotanshinone ameliorates radiationinduced lung injury in rats. Evidence-Based Complementary and Alternative Medicine. 2019; doi: 10.1155/2019/1908416.
- 46. Kabel AM. Free radicals and antioxidants: Role of enzymes and nutrition. World Journal of

- 47. Palace VP, Khaper N, Qin Q, Singal PK. Antioxidant potentials of vitamin A and carotenoids and their relevance to heart disease. Free Radic Biol Med. 1999; 26(5-6):746-61. doi: 10.1016/s0891-5849(98)00266-4.
- Tesoriere L, Ciaccio M, Bongiorno A, Riccio A, Pintaudi AM, Livrea MA. Antioxidant activity of all-trans-retinol in homogeneous solution and in phosphatidylcholine liposomes. Archives of biochemistry and biophysics. 1993; 307(1):217-23. doi: 10.1006/abbi.1993.1581.
- 49. Harapanhalli RS, Narra VR, Yaghmai V, Yaghmai V, Azure MT, Goddu SM, et al. Vitamins as radioprotectors in vivo II. Protection by vitamin A and soybean oil against radiation damage caused by internal radionuclides. Radiat Res. 1994; 139(1):115-22. PMID: 8016300.

- Massy ZA, Drücke TB. Magnesium and outcomes in patients with chronic kidney disease: Focus on vascular calcification, atherosclerosis and survival. Clin Kidney J. 2012; 5(Suppl 1):52-61. doi: 10.1093/ndtplus/sfr167.
- 51. Ryan CM, Geckle M. Why is learning and memory dysfunction in Type 2 diabetes limited to older adults?. Diabetes Metab Res Rev. 2000; 16(5):308-15. doi: 10.1002/1520-7560(2000)9999:9999<:::aiddmr141>3.0.co;2-x.
- 52. Yarube IU. Nitrate-induced oxidative stress and the effects of dietary antioxidant vitamins C, E and A: Insights from experimental and clinical studies. Bayero Journal of Pure and Applied Sciences. 2011; 4(2):69-79. doi: 10.4314/bajopas.v4i2.14.
- 53. Geiger H, Wanner C. Magnesium in disease. Clin kidney j. 2012; 5(1):25-38. doi: 10.1093/ndtplus/sfr165.