

Journal of Kerman University of Medical Sciences

Review Article





The Effect of Saffron and its Active Compounds on Oxidative Stress Markers in Diabetic Rats: A Systematic Review and Meta-analysis of Animal Studies

Yaser Mohammadi¹[®], Azam Rezaei Farimani²[®], Hossein Beydokhti³[®], Sameep Shetty⁴, Seyed Mohammad Riahi⁵[®]

¹Department of Biochemistry, School of Medicine, Iran University of Medical Sciences, Tehran, Iran

²Department of Clinical Biochemistry, School of Medicine, Cardiovascular Diseases Research Center, Birjand University of Medical Sciences, Birjand, Iran

³Department of General Courses, School of Medicine, Birjand University of Medical Sciences, Birjand, Iran ⁴Department of Oral and Maxillofacial Surgery, Manipal College of Dental Sciences, Mangalore, Manipal Academy of

Higher Education, A Constituent of MAHE, India

⁵Department of Community Medicine, Cardiovascular Diseases Research Center, Birjand University of Medical Sciences, Birjand, Iran

*Corresponding Author: Seyed Mohammad Riahi, Email: Riahim61@gmail.com

Abstract

Background: Diabetes, as a chronic metabolic disease, can induce oxidative stress, leading to severe damage to various tissues, including the kidneys, heart, and others. This study aimed to assess the influence of saffron and its active component on oxidative stress markers in diabetic rats.

Methods: The databases were searched until December 24, 2021. The quality of the included articles was assessed using SYRCLE's Risk of Bias tool. To estimate the effects of saffron and its active component, SMD with 95% confidence intervals (CIs) were pooled using a random-effects model. Subgroup analysis and meta-regression were used to explore heterogeneity. Publication bias was assessed using the Begg and Egger tests. The results were reported under the PRISMA guidelines.

Results: The meta-analysis comprising 42 articles revealed that prolonged hyperglycemia leads to increased oxidative markers, including malondialdehyde (MDA), nitric oxide (NO), total oxidant status (TOS), xanthine oxidase (XO), and reactive oxygen species (ROS)), and decreased antioxidant defense system, including glutathione (GSH), catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx), total antioxidant status (TAS), thiol groups (SH), and total antioxidant capacity (TAC). Treatment of diabetic rats with saffron, crocin, and safranal decreased the oxidant markers and increased the antioxidant markers. **Conclusion:** Saffron, crocin, and safranal reduce oxidative stress by reinforcing the antioxidant defense system and reducing oxidant markers. Hence, we believe that saffron and its active ingredients can be favorable options for managing diabetes and its complications. However, further human studies are required to draw definite conclusions.

Keywords: Diabetes, Oxidative stress, Saffron, Crocin, Safranal, Meta-analysis, Diabetic rats

Citation: Mohammadi Y, Rezaei Farimani A, Beydokhti H, Shetty S, Riahi SM. The effect of saffron and its active compounds on oxidative stress markers in diabetic rats: a systematic review and meta-analysis of animal studies. *Journal of Kerman University of Medical Sciences*. 2024;31(3):214–225. doi: 10.34172/jkmu.2024.34

Received: June 6, 2022, Accepted: September 2, 2023, ePublished: August 24, 2024

Introduction

Diabetes mellitus (DM) is a globally recognized noncommunicable chronic disease gaining epidemic proportions. The deficiency of insulin or the body's poor performance in effectively utilizing insulin can be due to auto-immune destruction of pancreatic cells, genetics, urbanization, and a sedentary lifestyle. The disease is affecting individuals throughout the age spectrum, and those who have it are estimated to reach 552 million by 2030 (1).

The loss of pancreatic insulin-producing cells (type 1 diabetes) or a reduction in insulin sensitivity in its target receptors in muscles and fatty tissues (type 2 diabetes) (2)

elucidates its pathophysiology. Chronic and uncontrolled hyperglycemia activates and initiates signals that cause severe structural damage, resulting in functional defects of various tissues such as the kidney, liver, pancreas, brain, and eyes. In due course, it can cause nephropathy, retinopathy, neuropathy, and micro/macrovascular tissue damage (3). The normal metabolism of substrates diverges to atypical pathways in the diabetic state, releasing toxic byproducts, such as reactive oxygen species (ROS).

The ROS released in uncontrolled hyperglycemia is triggered by protein kinase C activation, glucose autoxidation, oxidative phosphorylation, methylglyoxal



© 2024 The Author(s); Published by Kerman University of Medical Sciences. This is an open-access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

generation, and sorbitol formation (4). Free radicals can overwhelm the body's natural antioxidant defense mechanism and cause oxidative stress (5). Oxidative stress is associated with protein inactivation, protein glycation, and impaired glutathione metabolism, and damages various cellular components (6,7). Superoxide dismutase (SOD), glutathione (GSH), glutathione peroxidase (GPx), and catalase (CAT) are some of the free radical scavenging mechanisms that help remove superoxide, hydrogen peroxide, and hydroxyl radicals (8). Apoptosis and tissue dysfunction are eventually induced by oxidative stress produced by an imbalance between the antioxidant defense system and free radicals (3,5). As a result, rebalancing the oxidant-antioxidant defense system is a significant concern in diabetes. Diabetes is treated with various medicines, although synthetic treatments have always caused adverse reactions and other side effects (9). In this sense, using effective natural remedies instead of chemical and pharmaceutical treatments is preferable.

Saffron (Crocus sativus L.) is a small but prominent species from the Iridaceae family, known in Iran for its coloring, seasoning, and aromatizing properties. It is a food additive used in traditional medicine to cure various illnesses, including depression, cognitive problems, seizures, and cancer. In addition, saffron has anticancer, antioxidant, and anti-inflammatory properties and can improve memory function and learning ability. The pharmacological activities of saffron are attributed to many of its active constituents, including crocin, picrocrocin, and safranal (10). Evidence suggests that saffron might be used to treat diabetes. Although the hypoglycemic effects of saffron have been explored, the antioxidant benefits of saffron have not been well investigated. Therefore, this study aimed to see how saffron and its active components affect oxidative stress markers in a diabetic rat model.

Methods

The current systematic review and meta-analysis used the *Cochrane Handbook for Systematic Reviews of Intervention criteria* to investigate the effect of saffron and its active components on oxidative stress markers in diabetes. The results were reported under the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) Guidelines (11).

PECOS

- Population: Rats with induced diabetes.
- Comparison: Untreated diabetic rats (control group).
- Outcome: Oxidants (including malondialdehyde [MDA], nitric oxide [NO], total oxidant status [TOS], xanthine oxidase [XO], reactive oxygen species [ROS]) and antioxidants (including GSH, CAT, SOD, GPx, total antioxidant status [TAS], thiol groups [SH], total antioxidant capacity [TAC]).
- Study designs: experimental animal studies

Data sources and search strategy

Two researchers (Y.M., H.B.) independently conducted a comprehensive electronic search using the five following databases: PubMed (n=89), Scopus (n=164), ProQuest (n=57), Web of Science (n=24), and Cochrane (n=80) until December 24, 2021, with no language restrictions. Database sources were searched using relevant keywords from MeSH and non-MeSH terms. The Boolean search terms (AND and OR) were used for the search strategy. In addition, the reference lists of the selected articles were manually checked (n=2). An automated Google Scholar database search was also done to ensure that the most recent papers on the subject were considered (Table 1).

Eligibility criteria

Studies were deemed acceptable if they met the following criteria: (a) animal studies, (b) investigations on the effects of saffron, crocin, and safranal on oxidative stress markers, and (c) studies on DM. Exclusion criteria include (a) randomized controlled trials (RCTs), (b) studies on other diseases and other parameters, (c) conference abstracts, reviews, editorials, protocols, letters, case reports, and commentaries, (d) studies with incomplete data, and (e) interventions by combining saffron and its constituents with other compounds or supplements.

Study selection and data extraction

Endnote Reference Manager X9 was used to store all identified papers from each database, eliminating duplicates using the Endnote function "Find duplicates." Two reviewers (Y.M. and A.R.F.) evaluated the remaining publications individually with 97% agreement to verify their eligibility for inclusion after duplicate records were removed. The screening process was divided into two parts. Initially, titles and abstracts were examined, and unrelated research was eliminated. The remaining papers were reviewed for eligibility in the second step using the full-text version. Throughout the research selection process, any disagreements were settled by face-toface conversation or consultation with a third reviewer (S.M.R.). Finally, all eligible studies were included in this study. Microsoft Excel software was used to create a special form for data mining. The information in the document included the following: first author, year of publication, country of study, quality of study, animal breed, sample size, duration of intervention, dose of intervention (mg/ kg/d), mean and standard deviation (SD) of findings before and after the intervention.

Quality assessment

Two independent authors (Y.M., A.R.F.) assessed the articles using SYRCLE's Risk of Bias (RoB) tool, a tool specially designed for animal studies (12). This tool can be used to detect selection, performance, detection, erosion, and report bias in the included studies. The

data in the articles were labeled as high category risk of bias (H), unclear risk of bias (U), or low risk of bias (L). A "No" decision indicates a high risk of bias. When the text description was insufficient to determine the danger of bias, the term "Unclear" was used; a "Yes" decision indicates a low risk of bias. Studies with quality scores below 60% were considered high-risk in terms of bias and were excluded. Two reviewers (Y.M. and A.R.F.) independently performed quality assessment, with 97% agreement. Differences in scoring were resolved through discussion between the authors and consultation with a third author (S.M.R.) if necessary.

Statistical analysis

Table 1. Search strategy

Stata software Version 14.0 was used to analyze the data. We assessed the metabolic effects of saffron and its active components using standardized mean difference

(SMD) with a 95% confidence interval (CI). A randomeffects model was used to estimate SMD (13). Q and I^2 statistics were used to determine heterogeneity. I^2 values above 0.6 were considered heterogeneous. Subgroup analysis and meta-regression approaches were used to address heterogeneity (14,15). The Egger regression test, a parametric method, was used to determine publication bias. Sensitivity analysis was also conducted to detect the influence of a single study on the overall estimate by eliminating one study and repeating the analysis. A P value of 0.1 was considered significant in the analyses, with less than ten studies.

Results

Search results

Figure 1 depicts the steps of the article selection process. In total, 414 studies were found in various databases,

| Databases | Search Strategy | Result |
|-------------------|--|--------|
| Scopus | (TITLE-ABS-KEY (saffron*) OR TITLE-ABS-KEY (safron*) OR TITLE-ABS-KEY (crocus*) OR TITLE-ABS-KEY (safrana*) OR TITLE-ABS-KEY (crocetin*) OR TITLE-ABS-KEY (picrocrocin*) OR TITLE-ABS-KEY (crocetin*)) AND (TITLE-ABS-KEY (diabet*) OR TITLE-ABS-KEY (crocetin*)) AND (TITLE-ABS-KEY (crocetin*)) AND (TITLE-ABS-KEY (crocetin*)) AND (TITLE-ABS-KEY (crocetin*)) OR TITLE-ABS-KEY (crocetin*)) OR TITLE-ABS | 164 |
| PubMed | ((Antioxidan*[Text Word] OR "oxidativ*"[Text Word] OR "oxidative stress*"[Text Word] OR "Oxidative marker*"[Text Word] OR "Enzymatic oxidativ*"[Text Word] OR "anti-oxidative marker*"[Text Word] OR Oxidant*[Text Word] OR "Free radical*"[Text Word] OR Freeradical*[Text Word] OR Malondialdehyd*[Text Word] OR "Superoxide dismutas* "[Text Word] OR Glutathion*[Text Word] OR "Non-enzymatic oxidative*"[Text Word] OR "anti-oxidative marker*"[Text Word] OR "Catalas*"[Text Word] OR "Selenoglutathione peroxidase*"[Text Word] OR "oxidative stress"[MeSH Terms] OR "Antioxidants"[Mesh] OR "Malondialdehyde"[Mesh] OR "Superoxide Dismutase"[Mesh] OR "Glutathione"[Mesh] OR "Catalase"[MeSH]) AND (Saffron*[Text Word] OR safron*[Text Word] OR "Crocus*"[Text Word] OR Safrana*[Text Word] OR crocetin*[Text Word] OR picrocrocin*[Text Word] OR crocus*"[Mesh] OR "safranal" [Supplementary Concept] OR "picrocrocin" [Supplementary Concept] OR "Crocus"] (Mesh] (Diabet*[Text Word] OR "Diabetic Nephropathies"[Mesh] OR "Diabetes Mellitus"[Mesh] OR "Diabetes Mellitus, Type 2"[Mesh]) | 89 |
| Proquest | ((ti(Saffron*) OR ab(Saffron*) OR su(Saffron*) OR ti(safron*) OR ab(safron*) OR su(safron*) OR ti(Crocus*) OR ab(Crocus*) OR su(Crocus*) OR ti(Saffrana*) OR su(Saffrana*) OR su(Safrana*) OR ti(crocetin*) OR ab(crocetin*) OR su(crocetin*) OR ti(picrocrocin*) OR ab(picrocrocin*) OR ti(picrocrocin*) OR ab(crocetin*) OR ab(crocetin*) OR su(picrocrocin*) OR ti(crocetin*) OR ab(crocetin*) OR su(crocetin*) OR su(picrocrocin*) OR ti(crocetin*) OR ab(crocetin*) OR su(crocetin*) OR su(picrocrocin*) OR ti(crocetin*) OR ab(crocetin*) OR su(crocetin*) OR su(picrocrocin*) OR ti(crocetin*) OR ti(crocetin*) OR su(picrocrocin*) OR ab(Diabet*) OR su(Diabet*) OR ti("Diabetic Nephropathies") OR ab("Diabetic Nephropathies") OR su("Diabetic Nephropathies") OR ab(Antioxidan*) OR su(Antioxidan*) OR ti(coxidativ*) OR ab(coxidativ*) OR su(coxidativ*) OR su(coxidative stress")) OR ab(("oxidative stress")) OR su(("oxidative stress")) OR ti("coxidative marker*") OR ab("Coxidative marker*") OR ab("Crocus*) OR su("coxidative marker*") OR ti("erazymatic oxidativ*") OR su("inti-oxidative marker*") OR ab("Crocus*) OR ab("free radical" OR "free radical") OR ti("free radical" OR "free radical") OR ti("free radical") OR ab(("free radical") OR ab(("free radical") OR ab(("free radical") OR ab(("free radical") OR ab(("superoxide dismutase")) OR su(("superoxide dismutase")) OR su("superoxide dismutase")) OR su("superoxide dismutase") OR su("Non-enzymatic oxidative*") OR su("superoxide dismutase") OR su("superoxide dismuta | 57 |
| Web of Science | TI = (Saffron*) OR AB = (Saffron*) OR AK = (Saffron*) OR TI = (safron*) OR AB = (safron*) OR AK = (safron*) OR TI = (Crocus*) OR AB = (Crocus*) OR AK = (Crocus*) OR TI = (Safrana*) OR AB = (Safrana*) OR AK = (Safrana*) OR TI = (crocetin*) OR AB = (crocetin*) OR AK = (crocetin*) OR TI = (picrocrocin*) OR AB = (picrocrocin*) OR AK = (picrocrocin*) OR TI = (crocin*) OR AB = (crocetin*) OR AK = (crocin*) AND TI = (Diabet*) OR AB = (Diabet*) OR AK = (Diabet*) OR TI = ("Diabetic Nephropathies") OR AB = ("Diabetic Nephropathies") OR AK = ("Diabetic Nephropathies") AND TI = (Antioxidan*) OR AB = (Antioxidan*) OR AK = (Antioxidan*) OR TI = (oxidativ*) OR AB = (oxidativ*) OR AK = (oxidativ*) OR TI = ("oxidative stress*") OR AB = ("oxidative stress*") OR AK = (moxidative stress*") OR AK = (moxidative stress*") OR AK = (moxidative marker*") OR AB = (moxidative marker*") OR AB = (moxidative*") OR AB = ("Enzymatic oxidativ*") OR AK = (moxidativ*") OR TI = (moxidative stress*") OR AB = (moxidative marker*") OR AB = (moxidativ*") OR AK = (moxidativ*") OR AK = (moxidativ*") OR AB = (moxidative marker*") OR AB = (moxidative marker*") OR AK = (moxidativ*") OR AK = (moxidativ*) OR AB = (moxidativ*") OR AK = (moxidative marker*") OR AB = (moxidativ*") OR AK = (moxidativ*") OR AK = (moxidativ*) OR AB = (moxidativ*") OR AK = (moxidative marker*") OR TI = (moxidativ*") OR AK = (moxidativ*") OR AK = (moxidativ*) OR AB = (moxidativ*) OR AK = (moxidativ*") OR TI = (moxidativ*") OR AK = (moxidativ*") OR AK = (moxidativ*) OR AB = (moxidati*) OR AK = (moxidativ*") OR TI = (moxidativ*") OR AK = (moxidativ*") OR AK = (moxidativ*) OR AB = (moxidati*) OR AK = (moxidativ*") OR AB = (moxidativ*") OR AK = (moxidativ*") OR AK = (moxidativ****) OR AB = (moxidativ************************************ | 80 |

Table 1. Continued.

| Databases | Search Strategy | | | | | | | | |
|-----------|-----------------|---|----|--|--|--|--|--|--|
| | #1 | (Saffron*):ti.ab.kw=276 | | | | | | | |
| | #2 | (safron*):ti.ab.kw=4 | | | | | | | |
| | #3 | (Crocus*):ti,ab,kw = 145 | | | | | | | |
| | #4 | (Safrana*):ti,ab,kw=4 | | | | | | | |
| | #5 | (crocetin*):ti,ab,kw=28 | | | | | | | |
| | #6 | (picrocrocin*):ti,ab,kw=0 | | | | | | | |
| | #7 | (crocin*):ti,ab,kw=75 | | | | | | | |
| | #8 | MeSH descriptor: [Crocus] explode all trees=55 | | | | | | | |
| | #9 | #1 OR #2 OR #3 OR #4 OR #5 OR #6 OR #7 OR #8=349 | | | | | | | |
| | #10 | (Diabet*):ti,ab,kw=90985 | | | | | | | |
| | #11 | MeSH descriptor: [Diabetic Nephropathies] explode all trees=1412 | | | | | | | |
| | #12 | MeSH descriptor: [Diabetes Mellitus] explode all trees=30783 | | | | | | | |
| | #13 | MeSH descriptor: [Diabetes Mellitus, Type 2] explode all trees=17348 | | | | | | | |
| | #14 | #10 OR #11 OR #12 OR #13=91206 | | | | | | | |
| | #15 | (Antioxidan*):ti,ab,kw=12679 | | | | | | | |
| | #16 | (oxidativ*):ti,ab,kw=12436 | | | | | | | |
| | #17 | (oxidative stress*):ti,ab,kw=10239 | | | | | | | |
| | #18 | (Oxidative marker*):ti,ab,kw=3891 | | | | | | | |
| | #19 | (Enzymatic oxidativ*):ti,ab,kw=214 | | | | | | | |
| Cochrane | #20 | (anti-oxidative marker*):ti,ab,kw=69 | 24 | | | | | | |
| | #21 | (Oxidant*):ti,ab,kw=2091 | | | | | | | |
| | #22 | (Free radical*):ti,ab,kw=4533 | | | | | | | |
| | #23 | (Freeradical*):ti,ab,kw=145 | | | | | | | |
| | #24 | (Malondialdehyd*):ti,ab,kw=2998 | | | | | | | |
| | #25 | (Superoxide dismutas*):ti,ab,kw=2198 | | | | | | | |
| | #26 | (Glutathion*):ti,ab,kw = 3559 | | | | | | | |
| | #27 | (Non-enzymatic oxidative*):ti,ab,kw=52 | | | | | | | |
| | #28 | (anti-oxidative marker*):ti,ab,kw=69 | | | | | | | |
| | #29 | (Catalas*):ti,ab,kw=926 | | | | | | | |
| | #30 | (Selenoglutathione peroxidase*):ti,ab,kw=1 | | | | | | | |
| | #31 | MeSH descriptor: [Oxidative Stress] explode all trees=2978 | | | | | | | |
| | #32 | MeSH descriptor: [Antioxidants] explode all trees = 4812 | | | | | | | |
| | #33 | MeSH descriptor: [Malondialdehyde] explode all trees=1252 | | | | | | | |
| | #34 | MeSH descriptor: [Superoxide Dismutase] explode all trees = 774 | | | | | | | |
| | #35 | MeSH descriptor: [Glutathione] explode all trees=671 | | | | | | | |
| | #36 | MeSH descriptor: [Catalase] explode all trees=260 | | | | | | | |
| | #37 | #15 OR #16 OR #17 OR #18 OR #19 OR #20 OR #21 OR #22 OR #23 OR #24 OR #25 OR #26 OR #27 OR #28 OR #29 | | | | | | | |
| | OR #30 O | R #31 OR #32 OR #33 OR #34 OR #35 OR #36=25778 | | | | | | | |
| | #38 | #9 AND #14 AND #37=24 | | | | | | | |

and two were found in other sources. After removing duplicates (n = 228), the titles and abstracts of 188 articles were assessed, 78 of which were excluded from the study. A total of 110 full-text articles were reviewed, and 68 studies were omitted, leaving 42 papers to be included and evaluated for quality in the present study.

Study characteristics

The specifications of 42 eligible articles are shown in Table 2. These studies were done in Iran (n=26), China (n=2), Egypt (n=3), Greece (n=2), Turkey (n=8), and Poland (n=1) and were published between 2009 and 2021. They included 41 studies on rats and one study on mice. In the trials, 37 publications used streptozotocin (STZ) (dose range 30 to 75 mg/kg), two papers used a high-fat diet, and three articles used a combination of STZ and a high-fat diet to induce diabetes. The duration of the intervention using saffron, crocin, and safranal was from 14 to 70 days. The chemical used was crocin in 28 articles (dose range 2.5 to 150 mg/kg), saffron in 10 articles (dose range 0.25 to 5 mg/kg).

Quality assessment

Table 2 shows the results, and Figure S1 (Supplementary file 1) shows the details of the quality assessment of the included articles. The quality evaluation results showed that 15 studies scored 88.89, 21 articles scored 77.78, and 6 papers scored 66.67.

Meta-analysis results

In these studies, diabetic groups treated with saffron, crocin, and safranal were compared to diabetic control groups. The meta-analysis results are presented in Table 3 (subgroup analysis in different specimens) and Table 4 (subgroup analysis by type of intervention).

The subgroup analysis for the effect of saffron and its active compounds on antioxidant markers in different specimens The results of our meta-analysis showed that in the treated diabetic group compared with the diabetic control group, antioxidant markers, including GSH, CAT, SOD, GPx, TAS, TAC, and SH, increased with high effectiveness in the different specimens. However, the heterogeneity of the studies was high. Most studies had been performed on



Figure 1. Flowchart of the study selection process for the systematic review and meta-analysis

GSH (n=22), and the results showed that the effectiveness of saffron and its active compounds on GSH was more in the lung tissue and less in the brain tissue (for lung: SMD, 5.31 (µmol/L) [95% CI, 3.03 to 7.59]; P=0.001, I^2 =67.5%; for the brain: SMD, 0.79 (µmol/L) [95% CI, 0.32 to 1.25]; P=0.001, I^2 =0.9%).

The subgroup analysis for the effect of saffron and its active compounds on oxidant markers in different specimens The results of this study highlighted the effectiveness of saffron in decreasing the oxidant markers, including MDA, NO, TOS, XO, and ROS, in different specimens in the diabetic group compared to the control group.

MDA, an indicator of lipid peroxidation, is often used as a marker, and its level has been explored in several studies. The results showed that the highest effect of saffron and its active compounds on MDA was in the kidney tissue and its lowest effect was seen in the gastric mucosa (for kidney: SMD, -3.69 (µmol/L) [95% CI, -5.17 to -2.75]; P=0.001, $I^2=78.6\%$; for gastric mucosa: SMD, -1.41 Table 2. Characteristics of articles included in the meta-analysis

| Reference | Year | Country | Animal | Sample size in each group | Type of Diabetes induction/dosage (mg/kg) | Treatment period (day) | Type of intervention/ dosage (mg/kg) | Measurement* | Quality score |
|----------------------------|------|---------|--------|------------------------------------|---|------------------------------|--|-------------------------|------------------|
| Yaribeygi et al (16) | 2019 | Iran | Rats | 6 | STZ/45 | 56 | Crocin/40 | MDA, GSH, SOD, CAT | 88.89 |
| Yaribeygi et al (17) | 2019 | Iran | Rats | 6 | STZ/40 | 56 | Crocin/40 | MDA, GSH, SOD, CAT | 88.89 |
| Yaribeygi et al (18) | 2017 | Iran | Rats | 6 | STZ/40 | 56 | Crocin/40 | MDA | 88.89 |
| Wu et al (19) | 2018 | China | Rats | 8 | STZ/60 | 14 | Crocin/60 | MDA, SOD, NO | 88.89 |
| Talebanzadeh et al (20) | 2018 | Iran | Rats | 8 | STZ/60 | 35 | Saffron/200 | MDA, GPx, SOD, CAT | 77.78 |
| Sefidgar et al (21) | 2019 | Iran | Rats | 6 | STZ/60 | 28 | Crocin/60 | TAS, TOS | 88.89 |
| Samarghandian et al (22) | 2013 | Iran | Rats | 8 | STZ/60 | 28 | Safranal/0.25, 0.5, 0.75 | MDA, GSH, SOD, CAT, NO | 77.78 |
| Samarghandian et al (23) | 2014 | Iran | Rats | 10 | STZ/60 | 28 | Saffron/20, 40, 80 | GSH, SOD, CAT | 88.89 |
| Samarghandian et al (24) | 2016 | Iran | Rats | 9 | STZ/60 | 28 | Saffron/10, 20, 40 | MDA, GSH, SOD, CAT, NO | 88.89 |
| Samarghandian et al (25) | 2016 | Iran | Rats | 9 | STZ/60 | 28 | Crocin/10, 20, 30 | MDA, GSH, SOD, CAT, NO | 77.78 |
| Samarghandian et al (26) | 2013 | Iran | Rats | 8 | STZ/60 | 28 | Safranal/0.25, 0.5, 0.75 | MDA, GSH, SOD, CAT, NO | 88.89 |
| Samaha et al (27) | 2019 | Egypt | Rats | 10 | STZ/50 | 28 | Crocin/10 | MDA, GSH, SOD, TAC, CAT | 66.67 |
| Rajaei et al (28) | 2012 | Iran | Rats | 7 | STZ/55 | 42 | Crocin/15, 30, 60 | MDA, SH | 77.78 |
| Rahbani et al (29) | 2012 | Iran | Rats | 10 | STZ/75 | 56 | Saffron/40 | MDA, GSH, GPx, SOD, CAT | 66.67 |
| Qiuet al (30) | 2020 | China | Mice | 12 | High- fat diet | 56 | Crocin/50 | MDA, GSH, SOD, CAT, ROS | 77.78 |
| Motamedrad et al (31) | 2019 | Iran | Rats | 7 | STZ/60 | 21 | Saffron/25, 100 | MDA, TAC | 66.67 |
| Rahbani et al (32) | 2011 | Iran | Rats | 10 | STZ/65 | 56 | Saffron/40 | MDA, GSH, SOD, CAT | 88.89 |
| Margaritis et al (33) | 2020 | Greece | Rats | 6 | STZ/65 | 28 | Crocin/20, 50 | gsh, sod | 88.89 |
| Kianbakht et al (34) | 2009 | Iran | Rats | 10 | STZ/50 | 35 | Saffron (25, 100, 250) /Crocin (2.5, 5, 10)/ Safranal (0.25, 2, 5) | MDA, GSH | 77.78 |
| Hazman et al (35) | 2014 | Turkey | Rats | 8 | STZ (30)/High -fat diet | 28 | Safranal | TAS, TOS | 77.78 |
| Hazman et al (36) | 2015 | Turkey | Rats | 8 | STZ (30)/High -fat diet | 28 | Safranal | GSH, TAS, TOS | 77.78 |
| Hazman et al (37) | 2016 | Turkey | Rats | 8 | STZ (30)/High-fat diet | 42 | Crocin/150 | TAS, TOS | 77.78 |
| Hasanpour et al (38) | 2018 | Iran | Rats | 8 | STZ/65 | 35 | Saffron/200 | MDA, GPx, SOD, CAT | 88.89 |
| Yaribeygi et al (39) | 2018 | Iran | Rats | 6 | STZ/45 | 56 | Crocin/40 | MDA, GSH, SOD, CAT | 77.78 |
| Ghorbanzadeh et al (40) | 2016 | Iran | Rats | 7 | STZ/35 | 56 | Crocin/50 | MDA, GPx, SOD, CAT | 88.89 |
| Farshid et al (41) | 2016 | Iran | Rats | 8 | STZ/50 | 56 | Crocin/5, 10, 20 | MDA, SOD | 77.78 |
| Farshid et al (42) | 2015 | Iran | Rats | 6 | STZ/60 | 56 | Safranal/1 | MDA | 66.67 |
| El-Fawal et al (43) | 2018 | Egypt | Rats | 8 | High-fat diet | 70 | Crocin/50 | MDA | 77.78 |
| Bajerska et al (44) | 2013 | Poland | Rats | 6 | STZ/40 | 35 | Saffron | MDA, TAC | 88.89 |
| Bahmani et al (45) | 2016 | Iran | Rats | 10 | STZ/65 | 56 | Crocin/100 | GSH, SOD, TAC, CAT | 66.67 |
| Asri_Rezaei et al (46) | 2014 | Iran | Rats | 8 | STZ/50 | 42 | Crocin/12.5, 25, 50 | MDA, TAC | 77.78 |
| Ashrafi et al (47) | 2016 | Iran | Rats | 7 | STZ/60 | 35 | Crocin/200 | MDA, GPx, SOD, CAT | 88.89 |
| Altinoz et al (48) | 2015 | Turkey | Rats | 10 | STZ/50 | 21 | Crocin/20 | MDA, GSH | 77.78 |
| Altinoz et al (49) | 2014 | Turkey | Rats | 10 | STZ/50 | 21 | Crocin/20 | MDA, GSH, XO | 77.78 |
| Altinoz et al (50) | 2015 | Turkey | Rats | 10 | STZ/50 | 21 | Crocin/20 | MDA, GSH, XO | 77.78 |
| Ahmadi et al (51) | 2017 | Iran | Rats | 7 | STZ/55 | 42 | Crocin/15 | MDA, SH | 66.67 |
| AbouHany et al (52) | 2018 | Egypt | Rats | 10 | STZ/50 | 56 | Crocin/20 | MDA, GSH, SOD, NO | 77.78 |
| Tamaddonfard, E et al (53) | 2012 | Iran | Rats | 6 | STZ/60 | 30 | Crocin/7.5, 15, 30 | MDA, TAC | 77.78 |
| Altinoz et al (54) | 2014 | Turkey | Rats | 10 | STZ/50 | 21 | Crocin/20 | MDA, GSH, XO | 77.78 |
| Ayşegül et al (55) | 2020 | Turkey | Rats | 7 | STZ/65 | 21 | Crocin/50 | MDA, SOD, TAS, TOS | 88.89 |
| Kapucu et al (56) | 2020 | Greece | Rats | 5 | STZ/55 | 70 | Saffron/60 | GPx, CAT | 77.78 |
| Yaribeygi et al (57) | 2021 | Iran | Rats | 6 | STZ/45 | 56 | Crocin/40 | MDA, GSH, SOD, CAT | 77.78 |

Abbreviations: glutathione (GSH), catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx), total antioxidant status (TAS), total antioxidant capacity (TAC), malondialdehyde (MDA), nitric oxide (NO), total oxidant status (TOS), thiol groups (SH), xanthine oxidase (XO), reactive oxygen species (ROS).

| Table 3 | Effect | of sa | affron | and | its | effective | compo | ounds | on | oxidative | stress |
|---------|----------------------------|--------|---------|-------|------|-------------|----------|--------|------|-----------|--------|
| markers | (oxidar | nts ar | nd anti | oxida | ants | s) accordii | ng to th | e type | e of | speciment | 5 |

Table 3. Continued.

| Variable | N | SMD (95% CI) | P value for SMD | Q | ľ | Tau |
|-------------------|----|---------------------|--------------------|-------|------|--------|
| | | Antioxidant | markers | | | |
| GSH | | | | | | |
| Gastric mucosa | 9 | 4.94 (4, 5.88) | < 0.001 | 18.56 | 0.0 | 0.00 |
| Liver | 6 | 0.87 (0.44, 1.29) | < 0.001 | 1.98 | 0.0 | 0.00 |
| BALF | 3 | 3.87 (2.71, 5.03) | < 0.001 | 2.58 | 22.6 | 0.24 |
| Lung | 3 | 5.31 (3.03, 7.59) | < 0.001 | 6.14 | 67.5 | 2.72 |
| Serum | 11 | 1.06 (0.75, 1.36) | < 0.001 | 8.98 | 0.0 | 0.00 |
| Testis | 1 | 2.96 (1.24, 4.68) | < 0.001 | 0.00 | 0.0 | 0.00 |
| Brain | 4 | 0.79 (0.32, 1.25) | < 0.001 | 3.03 | 0.9 | 0.0021 |
| Heart | 1 | 4.99 (3.14, 6.85) | < 0.001 | 0.00 | 0.0 | 0.00 |
| Kidney | 7 | 1.09 (-0.37, 2.55) | 0.145 | 57.30 | 89.5 | 3.33 |
| Lens | 1 | 4.43 (2.74, 6.12) | < 0.001 | 0.00 | 0.0 | 0.00 |
| Pancreas | 2 | 3.82 (-2.99, 10.63) | 0.272 | 13.87 | 92.8 | 22.46 |
| CAT | | | | | | |
| Liver | 3 | 1.49 (-1.15, 4.14) | 0.26 | 26.37 | 92.4 | 4.84 |
| BALF | 3 | 3.65 (1.71, 5.59) | < 0.001 | 7.64 | 73.8 | 2.13 |
| Lung | 3 | 2.67 (1.6, 3.74) | < 0.001 | 3.39 | 40.9 | 0.36 |
| Serum | 10 | 0.93 (0.62, 1.24) | < 0.001 | 5.32 | 0.0 | 0.00 |
| Brain | 3 | 0.74 (0.16, 1.31) | 0.012 | 2.35 | 14.8 | 0.038 |
| Lens | 2 | 2.76 (1.82, 3.71) | < 0.001 | 0.04 | 0.0 | 0.00 |
| Kidney | 2 | 4.59 (1.28, 7.9) | 0.007 | 3.87 | 74.1 | 4.35 |
| Heart | 1 | 4.66 (2.52, 6.80) | < 0.001 | 0.00 | 0.0 | 0.00 |
| Testis | 2 | 5.37 (-3.11, 13.86) | 0.215 | 13.33 | 92.5 | 34.81 |
| Pancreas | 2 | 1.39 (0.47, 2.30) | 0.003 | 0.34 | 0.0 | 0.00 |
| Retina | 1 | 4.83 (2.16, 7.50) | < 0.001 | 0.00 | 0.0 | 0.00 |
| SOD | | | | | | |
| Liver | 5 | 1.05 (-0.24, 2.35) | 0.11 | 22.68 | 82.4 | 1.74 |
| BALF | 3 | 3.06 (1.82, 4.31) | < 0.001 | 3.95 | 49.4 | 0.59 |
| Lung | 3 | 3.69 (1.55, 5.84) | < 0.001 | 9.32 | 78.5 | 2.71 |
| Serum | 11 | 1.88 (1.21, 2.55) | < 0.001 | 35.89 | 72.1 | 0.88 |
| Brain | 3 | 0.90 (0.32, 1.49) | < 0.003 | 2.36 | 15.1 | 0.04 |
| Lens | 2 | 1.77 (-2.28, 5.82) | 0.392 | 17.14 | 94.2 | 8.04 |
| Heart | 4 | 4.90 (3.81, 6) | < 0.001 | 3.14 | 4.5 | 0.05 |
| Kidney | 6 | 1.07 (-1.86, 4) | 0.473 | 91.94 | 94.6 | 12.51 |
| Testis | 2 | 4.12 (-4.79, 13.02) | 0.365 | 18.25 | 94.5 | 39.12 |
| Pancreatic | 2 | 2.66 (-0.65, 5.97) | 0.116 | 6.62 | 84.9 | 4.87 |
| GPx | | | | | | |
| Liver | 1 | 0.80 (-0.11, 1.72) | 0.085 | 0.00 | 0 | 0.00 |
| Lens | 1 | 2.42 (1.09, 3.75) | < 0.001 | 0.00 | 0.0 | 0.00 |
| Testis | 1 | 2.40 (1.08, 3.73) | < 0.001 | 0.00 | 0.0 | 0.00 |
| Heart | 1 | 4.30 (2.29, 6.32) | < 0.001 | 0.00 | 0.0 | 0.00 |
| Kidney | 1 | 4.81 (2.90, 6.32) | < 0.001 | 0.00 | 0.0 | 0.00 |

| Variable | Ν | SMD (95% CI) | P value for SMD | Q | I ² | Tau |
|-------------------|----|----------------------|-----------------|-------|-----------------------|-------|
| TAS | | | | | | |
| Serum | 6 | 0.84 (-0.32, 2.01) | 0.15 | 29.24 | 82.9 | 1.74 |
| Pancreas | 4 | 0.14 (-1.38, 1.10) | 0.82 | 16.13 | 81.4 | 1.29 |
| Kidney | 3 | 2.40 (0.12, 4.68) | 0.03 | 15.52 | 87.1 | 3.37 |
| TAC | | | | | | |
| Serum | 10 | 2.29 (1.36, 3.21) | < 0.001 | 32.63 | 72.4 | 1.53 |
| SH | | | | | | |
| Liver | 3 | 0.78 (0.14, 1.41) | 0.016 | 0.24 | 0.0 | 0.00 |
| Kidney | 3 | 1.68 (0.95-2.40) | < 0.001 | 1.37 | 0.0 | 0.00 |
| Brain | 3 | 0.17 (-0.65, 0.98) | 0.68 | 3.41 | 41.4 | 0.21 |
| | | Oxidant m | arkers | | | |
| MDA | | | | | | |
| Liver | 7 | -1.61 (-2.17, -1.06) | < 0.001 | 9.27 | 35.3 | 0.19 |
| Serum | 22 | -1.87 (-2.36, -1.38) | < 0.001 | 69.41 | 69.7 | 0.90 |
| Kidney | 10 | -3.69 (-5.17, -2.75) | < 0.001 | 42 | 78.6 | 2.78 |
| Heart | 5 | -3.44 (-4.94, -1.94) | < 0.001 | 17.76 | 77.5 | 2.18 |
| Brain | 3 | -1.47 (-2.18, -0.75) | < 0.001 | 4.54 | 33.9 | 0.17 |
| Pancreas | 2 | -1.71 (-3.06, -0.36) | 0.013 | 1.85 | 46.1 | 0.44 |
| BALF | 3 | -1.95 (-3.12, -0.78) | < 0.001 | 5.20 | 61.6 | 0.65 |
| Lung | 3 | -2.41 (-3.57, -1.25) | < 0.001 | 4.35 | 54 | 1.8 |
| Gastric mucosa | 9 | -1.41 (-2.23, -0.59) | < 0.001 | 44.73 | 82.1 | 0.00 |
| Testis | 2 | -3.38 (-10.91, 4.15) | 0.379 | 17.76 | 94.4 | 27.92 |
| Sciatic nerve | 2 | -2.69 (-3.85, -1.53) | < 0.001 | 0.39 | 0 | 0.00 |
| NO | | | | | | |
| BALF | 3 | -2.47 (-4.07, -0.88) | 0.002 | 7.93 | 74.8 | 1.46 |
| Lung | 3 | -4.67 (-7.63, -1.71) | 0.002 | 12.84 | 84.4 | 5.65 |
| Serum | 9 | -2.13 (-2.95, -1.31) | < 0.001 | 31.54 | 74.6 | 1.12 |
| Kidney | 1 | -4.67 (-6.43, -2.91) | < 0.001 | 0.00 | 0 | 0.00 |
| TOS | | | | | | |
| Serum | 6 | -1.50 (-3.38, 0.38) | 0.11 | 56.90 | 91.2 | 4.82 |
| Pancreas | 4 | -1.49 (-3.29, -0.32) | 0.10 | 26.18 | 88.5 | 2.96 |
| Kidney | 3 | -1.26 (-3.59, -1.06) | 0.287 | 20.51 | 90.2 | 3.76 |
| XO | | | | | | |
| Liver | 1 | -1.40 (-2.39, -0.41) | < 0.001 | 0.00 | 0 | 0.00 |
| Brain | 1 | -1.60 (-2.62, -0.58) | < 0.001 | 0.00 | 0 | 0.00 |
| Kidney | 1 | -1.44 (-2.44, -0.45) | < 0.001 | 0.00 | 0 | 0.00 |
| ROS | | | | | | |
| Serum | 1 | -0.80 (-1.63, 0.04) | < 0.001 | 0.00 | 0 | 0.00 |

Abbreviations: glutathione (GSH), catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx), total antioxidant status (TAS), total antioxidant capacity (TAC), malondialdehyde (MDA), nitric oxide (NO), total oxidant status (TOS), thiol groups (SH), xanthine oxidase (XO), reactive oxygen species (ROS).

 Table 4. Comparison of oxidative stress markers (oxidants and antioxidants)

 by type of intervention

Antioxidant markers

SMD (95%CI)

1.58 (0.92, 2.24)

2.16 (1.45, 2.86)

3.28 (1.97, 4.6)

Variable N

11

23

14

GSH

Saffron

Crocin

Safranal

CAT

P value

for SMD

< 0.001

< 0.001

< 0.001

12

76.4

86

90.7

Tau

0.92

2.4

5.53

0

42.38

157.64

140.14

| Effect of saffron on oxidative str | ess |
|------------------------------------|-----|
|------------------------------------|-----|

(μ mol/L) [95% CI, -2.23 to -0.59]; P = 0.001, $I^2 = 82.1\%$).

The subgroup analysis for comparison of oxidative stress markers (oxidants and antioxidants) by type of intervention The results of our meta-analysis showed that saffron, crocin, and safranal increase the levels of antioxidants markers (GSH, CAT, SOD, GPx, TAS, TAC, and SH) and decrease the levels of oxidant markers (MDA, NO, TOS, XO, and ROS). The results of our meta-analysis suggest that safranal was more effective than saffron and crocin on antioxidant and oxidant markers. In addition, saffron showed the least effectiveness compared to crocin and safranal.

Meta-regression

Meta-regression was performed to evaluate the effect of quantitative covariates on each of the markers of oxidative stress (oxidant and antioxidant), intervention dose, duration of intervention, publication year, quality score, and sample size. Meta-regression for MDA showed that there was no relationship between intervention dose (b = -0.0061; [95% CI = -0.01 to 0.003]; P = 0.2), duration of the intervention (b = -0.0057; [95% CI = -0.03 to 0.02]; P=0.6), publication year (b=-0.16; [95% CI=-0.30 to -0.03]; P = 0.01), quality score (b = -0.011; [95% CI = -0.078to 0.05]; P=0.7), and sample size (b=0.22; [95%) CI = -0.059 to 0.51]; P = 0.1). Figure 2 shows the metaregression for MDA. This analysis was performed for other oxidative stress markers (oxidant and antioxidant), but no significant relationship was observed (data are not shown).

Publication bias

In this study, publication bias was assessed by funnel plot and Egger's test, and it was observed that there was no evidence of publication bias (*P*-value $_{Egger's test} > 0.05$) (data are not shown).

Sensitivity analysis

In this study, sensitivity analysis was performed by excluding studies with a quality score below 70%, and no change was observed in the results (data are not shown).

Discussion

Main findings

This is the first systematic review and meta-analysis to comprehensively investigate the effect of saffron and its active compounds on oxidative stress markers (oxidants and antioxidants).

DM is a complex metabolic disease caused by a defect in the amount or function of insulin (52). Various studies show that oxidative stress begins at the onset of diabetes and gradually increases following long-term hyperglycemia (44,53). Free radicals facilitate tissue damage through lipid peroxidation, changes in antioxidant enzyme activity,

| Saffron | 12 | 1.15 (0.52, 1.79) | < 0.001 | 45.80 | 76 | 0.91 |
|---------------|----|----------------------|---------|--------|------|------|
| Crocin | 11 | 2.35 (1.38, 3.31) | < 0.001 | 54.40 | 81.6 | 1.9 |
| Safranal | 9 | 2.48 (1.73, 3.23) | < 0.001 | 21.51 | 62.8 | 0.79 |
| SOD | | | | | | |
| Saffron | 12 | 1.57 (0.90, 2.24) | < 0.001 | 45.06 | 75.6 | 0.98 |
| Crocin | 21 | 2.01 (1.02, 3.01) | < 0.001 | 203.59 | 90.2 | 4.57 |
| Safranal | 9 | 3.33 (2.58, 4.08) | < 0.001 | 15.62 | 48.8 | 0.62 |
| GPx | | | | | | |
| Saffron | 5 | 3.14 (1.24, 5.04) | < 0.001 | 27.77 | 85.6 | 3.43 |
| Crocin | 1 | 4.30 (2.29, 6.32) | < 0.001 | 0.00 | 0 | 0.00 |
| TAS | | | | | | |
| Crocin | 7 | 1.06 (-0.18, 2.30) | 0.094 | 42.20 | 85.8 | 2.31 |
| Safranal | 6 | 0.60 (-0.55, 1.75) | 0.30 | 31.92 | 84.3 | 1.72 |
| TAC | | | | | | |
| Saffron | 3 | 3.90 (0.78, 7.03) | 0.01 | 15.26 | 86.9 | 6.41 |
| Crocin | 7 | 1.82 (0.98, 2.67) | < 0.001 | 14.5 | 58.6 | 0.74 |
| SH | | | | | | |
| Crocin | 9 | 0.84 (0.32, 1.35) | 0.001 | 14.63 | 45.3 | 0.27 |
| | | Oxidant | markers | | | |
| MDA | | | | | | |
| Saffron | 15 | -2.55 (-3.58, -1.53) | < 0.001 | 120.76 | 88.4 | 3.13 |
| Crocin | 43 | -2.20 (-2.58, -1.82) | < 0.001 | 146.73 | 71.4 | 1.09 |
| Safranal | 13 | -2.14 (-2.75,153) | < 0.001 | 36.11 | 66.8 | 0.81 |
| NO | | | | | | |
| Saffron | 3 | -1.58 (-2.72, -0.43) | 0.007 | 6.44 | 68.9 | 0.7 |
| Crocin | 5 | -2.18 (-3.35, -1.02) | < 0.001 | 18.13 | 77.9 | 1.34 |
| Safranal | 8 | -3.70 (-4.97, -2.43) | < 0.001 | 32.60 | 78.5 | 2.5 |
| TOS | | | | | | |
| Crocin | 7 | -1.44 (-2.70, -0.18) | 0.025 | 40.42 | 85.2 | 2.42 |
| Safranal | 6 | -1.40 (-3.19, 0.39) | 0.125 | 58.36 | 91.4 | 4.33 |
| ХО | | | | | | |
| | 3 | -1.48 (-2.06, -0.90) | 0.001 | 0.08 | 0.0 | 0.00 |
| Crocin | | | | | | |
| Crocin ROS | | | | | | |

total oxidant status (TOS), thiol groups (SH), xanthine oxidase (XO), reactive

oxygen species (ROS).



Figure 2. Meta-regression for malondialdehyde (MDA) based on a: Intervention dose, b: Duration of intervention, c: Publication year, d: Quality score, and f: Sample size

nuclear factor kappa B (NF- $\kappa\beta$) activation, and induction of apoptosis (47). MDA disrupts membrane function by reducing membrane fluidity and altering the activity of membrane-bound enzymes and receptors, eventually leading to damage to cellular components (22).

A mounting body of evidence in the literature has highlighted the hypoglycemic properties of saffron, making it a potential nutraceutical for diabetes. The enhancement in insulin sensitivity, initiation of insulin signaling pathways, enhancement of β -cell actions, advancement of glucose transporter type 4 (GLUT-4) expression, reduction of oxidative stress, and suppression of the expression of inflammatory mediators explains the hypoglycemic action of saffron (23, 58).

Our meta-analysis showed that saffron and its active compounds significantly decrease oxidant markers, including MDA, NO, TOS, XO, and ROS, in different tissues, with a large effect size. Although there is heterogeneity in some studies, numerous studies have shown that saffron, crocin, and safranal have a hypoglycemic effect (22,38,51). Thus, according to our results and previous studies, saffron and its active compounds have protective effects on different tissues by reducing oxidant markers and hyperglycemia.

The results of our meta-analysis showed that saffron and its active compounds increased antioxidant markers, including GSH, CAT, SOD, GPx, TAS, TAC, and SH, in different tissues, with a large effect size. SOD, CAT, and GSH are defense factors against free radicals (30). However, the heterogeneity of the studies was substantial. Long-term hyperglycemia decreases antioxidant enzyme function due to non-enzymatic glycation or structural damage caused by excessive ROS generation (59). SOD is a metalloprotein and the first enzyme in the antioxidant defense chain that converts O⁻² to H₂O₂ (24), followed by CAT and GPx. CAT is localized in peroxisomes or microperoxysomes, which catalyze the breakdown of H₂O₂ into water and oxygen (23,25). GPx uses GSH as a proton donor to convert H₂O₂ to H₂O and O₂ (56). Taking a cue from this metanalysis, saffron, crocin, and safranal reinforce the antioxidant defense system and protect different tissues in hyperglycemic conditions.

The results show that saffron and its active compounds (crocin and safranal) have an overall protective effect against oxidative stress. In our study, safranal showed a larger effect on oxidative stress markers than saffron and crocin. Among the studies included in the meta-analysis, the study of Kianbakht et al examined the concurrent effect of saffron, crocin, and safranal. It concluded that safranal is more effective than saffron and crocin in minimizing oxidative stress (34). Safranal is a monoterpene aldehyde and a principal constituent of the essential oil of saffron, but it should be noted that safranal is usually present in less than 1% of saffron (60). saffron contains more than 150 chemicals (61) and may be less effective than safranal due to its diverse chemicals. Therefore, further research on different saffron, crocin, and safranal doses is necessary to confirm the hypothesis.

Strengths and limitations

Strengths: An exploratory search was conducted to identify articles on the role of saffron and its active compounds in diabetes and oxidative stress markers, and the results were included in the meta-analysis. A subgroup analysis was done based on the type of specimen and intervention. Lastly, we tried to identify the causes of heterogeneity using meta-regression and subgroup analysis techniques.

Limitations: First, in a few of the analyses, the total number of included articles was low, especially papers on XO and ROS. Second, some studies showed high heterogeneity in the analyses.

Conclusion

The radical scavenging trait of saffron is well known to reinforce the antioxidant defense system. Saffron can play a role in offsetting the multifactor metabolic disorder seen in DM by modulating the glucose levels and the expression of inflammatory mediators. Also, further in vivo human studies are essential to confirm the affirmative effects of saffron as a complementary therapy for DM.

Acknowledgments

The authors would like to thank Birjand University of Medical Sciences.

Authors' Contribution

Conceptualization: Yaser Mohammadi, Seyed Mohammad Riahi, Azam Rezaei Farimani. Data curation: Seyed Mohammad Riahi, Hossein Beydokhti. Investigation: Seyed Mohammad Riahi, Yaser Mohammadi, Azam Rezaei Farimani, Azam Rezaei Farimani. Methodology: Seyed Mohammad Riahi, Hossein Beydokhti.

Project administration: Seyed Mohammad Riahi.

Resources: Seyed Mohammad Riahi.

Software: Seyed Mohammad Riahi, Hossein Beydokhti.

Supervision: Seyed Mohammad Riahi.

Validation: Sameep Shetty, Yaser Mohammadi, Seyed Mohammad Riahi, Azam Rezaei Farimani, Hossein Beydokhti.

Visualization: Seyed Mohammad Riahi. Writing-original draft: Yaser Mohammadi.

Writing-review & editing: Sameep Shetty, Seyed Mohammad Riahi, Yaser Mohammadi, Azam Rezaei Farimani, Hossein Beydokhti.

Competing Interests

The authors declare that they have no conflict of interest.

Ethical Approval

This study was approved by Birjand University of Medical Sciences Ethics Committee (IR.BUMS.REC.1400.138).

Funding

None.

Supplementary Files

Supplementary file 1 contains Figure S1.

References

- Shaw JE, Sicree RA, Zimmet PZ. Global estimates of the prevalence of diabetes for 2010 and 2030. Diabetes Res Clin Pract. 2010;87(1):4-14. doi: 10.1016/j.diabres.2009.10.007.
- Abdul-Hamid M, Moustafa N. Protective effect of curcumin on histopathology and ultrastructure of pancreas in the alloxan treated rats for induction of diabetes. J Basic Appl Zool. 2013;66(4):169-79. doi: 10.1016/j.jobaz.2013.07.003.
- Arora MK, Singh UK. Molecular mechanisms in the pathogenesis of diabetic nephropathy: an update. Vascul Pharmacol. 2013;58(4):259-71. doi: 10.1016/j. vph.2013.01.001.
- Rains JL, Jain SK. Oxidative stress, insulin signaling, and diabetes. Free Radic Biol Med. 2011;50(5):567-75. doi: 10.1016/j.freeradbiomed.2010.12.006.
- Zhang P, Li T, Wu X, Nice EC, Huang C, Zhang Y. Oxidative stress and diabetes: antioxidative strategies. Front Med. 2020;14(5):583-600. doi: 10.1007/s11684-019-0729-1.
- Kashihara N, Haruna Y, Kondeti VK, Kanwar YS. Oxidative stress in diabetic nephropathy. Curr Med Chem. 2010;17(34):4256-69. doi: 10.2174/092986710793348581.
- Yaribeygi H, Sathyapalan T, Atkin SL, Sahebkar A. Molecular mechanisms linking oxidative stress and diabetes mellitus. Oxid Med Cell Longev. 2020;2020:8609213. doi: 10.1155/2020/8609213.
- Yaribeygi H, Mohammadi MT, Sahebkar A. Crocin potentiates antioxidant defense system and improves oxidative damage in liver tissue in diabetic rats. Biomed Pharmacother. 2018;98:333-7. doi: 10.1016/j.biopha.2017.12.077.
- 9. Elgazar AF, Rezq AA, Bukhari HM. Anti-hyperglycemic effect

of saffron extract in alloxan-induced diabetic rats. Eur J Biol Sci. 2013;5(1):14-22.

- Cardone L, Castronuovo D, Perniola M, Cicco N, Candido V. Saffron (*Crocus sativus* L.), the king of spices: an overview. Sci Hortic. 2020;272:109560. doi: 10.1016/j. scienta.2020.109560.
- Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, et al. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. Syst Rev. 2015;4(1):1. doi: 10.1186/2046-4053-4-1.
- Hooijmans CR, Rovers MM, de Vries RB, Leenaars M, Ritskes-Hoitinga M, Langendam MW. SYRCLE's risk of bias tool for animal studies. BMC Med Res Methodol. 2014;14:43. doi: 10.1186/1471-2288-14-43.
- 13. Faraone SV. Interpreting estimates of treatment effects: implications for managed care. P T. 2008;33(12):700-11.
- Mokhayeri Y, Riahi SM, Rahimzadeh S, Pourhoseingholi MA, Hashemi-Nazari SS. Metabolic syndrome prevalence in the Iranian adult's general population and its trend: a systematic review and meta-analysis of observational studies. Diabetes Metab Syndr. 2018;12(3):441-53. doi: 10.1016/j. dsx.2017.12.023.
- 15. Riahi SM, Mokhayeri Y. Methodological issues in a metaanalysis. Curr Med Res Opin. 2017;33(10):1813. doi: 10.1080/03007995.2017.1359152.
- Yaribeygi H, Noroozadeh A, Mohammadi MT, Johnston TP, Sahebkar A. Crocin improves oxidative stress by potentiating intrinsic anti-oxidant defense systems in pancreatic cells during uncontrolled hyperglycemia. J Pharmacopuncture. 2019;22(2):83-9. doi: 10.3831/kpi.2019.22.010.
- 17. Yaribeygi H, Mohammadi MT, Sahebkar A. Crocin potentiates antioxidant defense system and improves oxidative damage in liver tissue in diabetic rats. Biomed Pharmacother. 2018;98:333-7. doi: 10.1016/j.biopha.2017.12.077.
- Yaribeygi H, Mohammadi MT. Protective Effect of Crocin on Kidney Performance in Chronic Uncontrolled Hyperglycemia-Induced Nephropathy in Rat. J Adv Med Biomed Res. 2017;25(109):36-49. [Persian].
- 19. Wu X, Long E, Wang L, Li G, Yang Y, Liu H, et al. Crocin enhances antioxidative and cardioprotective effects of sitagliptin in streptozotocin-induced diabetic rats. Int J Clin Exp Med. 2018;11(7):6848-55.
- Talebanzadeh S, Ashrafi M, Kazemipour N, Erjaee H, Nazifi S. Evaluation of the effects of saffron aqueous extract on oxidative stress in the lens of streptozotocin-induced diabetic rats. Biomed Res Ther. 2018;5(4):2133-41. doi: 10.15419/ bmrat.v5i4.427.
- Sefidgar SM, Ahmadi-Hamedani M, Jebelli Javan A, Narenji Sani R, Javaheri Vayghan A. Effect of crocin on biochemical parameters, oxidative/antioxidative profiles, sperm characteristics and testicular histopathology in streptozotocininduced diabetic rats. Avicenna J Phytomed. 2019;9(4):347-61.
- 22. Samarghandian S, Borji A, Delkhosh MB, Samini F. Safranal treatment improves hyperglycemia, hyperlipidemia and oxidative stress in streptozotocin-induced diabetic rats. J Pharm Pharm Sci. 2013;16(2):352-62. doi: 10.18433/j3zs3q.
- 23. Samarghandian S, Azimi-Nezhad M, Samini F. Ameliorative effect of saffron aqueous extract on hyperglycemia, hyperlipidemia, and oxidative stress on diabetic encephalopathy in streptozotocin induced experimental diabetes mellitus. Biomed Res Int. 2014;2014:920857. doi: 10.1155/2014/920857.
- Samarghandian S, Azimi-Nezhad M, Farkhondeh T. Immunomodulatory and antioxidant effects of saffron aqueous extract (*Crocus sativus* L.) on streptozotocin-induced diabetes in rats. Indian Heart J. 2017;69(2):151-9. doi: 10.1016/j.

ihj.2016.09.008.

- Samarghandian S, Azimi-Nezhad M, Farkhondeh T. Crocin attenuate tumor necrosis factor-alpha (TNF-α) and interleukin-6 (IL-6) in streptozotocin-induced diabetic rat aorta. Cytokine. 2016;88:20-8. doi: 10.1016/j.cyto.2016.08.002.
- Samarghandian S, Afshari R, Sadati A. Evaluation of lung and bronchoalveolar lavage fluid oxidative stress indices for assessing the preventing effects of safranal on respiratory distress in diabetic rats. ScientificWorldJournal. 2014;2014:251378. doi: 10.1155/2014/251378.
- 27. Samaha MM, Said E, Salem HA. A comparative study of the role of crocin and sitagliptin in attenuation of STZ-induced diabetes mellitus and the associated inflammatory and apoptotic changes in pancreatic β-islets. Environ Toxicol Pharmacol. 2019;72:103238. doi: 10.1016/j.etap.2019.103238.
- Rajaei Z, Hadjzadeh MA, Nemati H, Hosseini M, Ahmadi M, Shafiee S. Antihyperglycemic and antioxidant activity of crocin in streptozotocin-induced diabetic rats. J Med Food. 2013;16(3):206-10. doi: 10.1089/jmf.2012.2407.
- Rahbani M, Mohajeri D, Rezaie A, Nazeri M. Protective effect of ethanolic extract of saffron (dried stigmas of *Crocus sativus* L.) on hepatic tissue injury in streptozotocin-induced diabetic rats. J Anim Vet Adv. 2012;11(12):1985-94. doi: 10.3923/ javaa.2012.1985.1994.
- Qiu Y, Jiang X, Liu D, Deng Z, Hu W, Li Z, et al. The hypoglycemic and renal protection properties of crocin via oxidative stress-regulated NF-κB signaling in db/db mice. Front Pharmacol. 2020;11:541. doi: 10.3389/fphar.2020.00541.
- 31. Motamedrad M, Shokouhifar A, Hemmati M, Moossavi M. The regulatory effect of saffron stigma on the gene expression of the glucose metabolism key enzymes and stress proteins in streptozotocin-induced diabetic rats. Res Pharm Sci. 2019;14(3):255-62. doi: 10.4103/1735-5362.258494.
- Rahbani M, Mohajeri D, Rezaie A, Doustar Y, Nazeri M. Attenuation of oxidative stress of hepatic tissue by ethanolic extract of saffron (dried stigmas of *Crocus sativus* L.) in streptozotocin (STZ)-induced diabetic rats. Afr J Pharm Pharmacol. 2011;5(19):2166-73. doi: 10.5897/ajpp11.624.
- Margaritis I, Angelopoulou K, Lavrentiadou S, Mavrovouniotis IC, Tsantarliotou M, Taitzoglou I, et al. Effect of crocin on antioxidant gene expression, fibrinolytic parameters, redox status and blood biochemistry in nicotinamide-streptozotocininduced diabetic rats. J Biol Res (Thessalon). 2020;27:4. doi: 10.1186/s40709-020-00114-5.
- Kianbakht S, Mozafari K. Effects of saffron and its active constituents, crocin and safranal, on prevention of indomethacin induced gastric ulcers in diabetic and nondiabetic rats. J Med Plants. 2009;8(Suppl 5):30-8.
- Hazman Ö, Ovalı S. Investigation of the anti-inflammatory effects of safranal on high-fat diet and multiple lowdose streptozotocin induced type 2 diabetes rat model. Inflammation. 2015;38(3):1012-9. doi: 10.1007/s10753-014-0065-1.
- Hazman Ö, Bozkurt MF. Anti-inflammatory and antioxidative activities of safranal in the reduction of renal dysfunction and damage that occur in diabetic nephropathy. Inflammation. 2015;38(4):1537-45. doi: 10.1007/s10753-015-0128-y.
- Hazman Ö, Aksoy L, Büyükben A. Effects of crocin on experimental obesity and type-2 diabetes. Turk J Med Sci. 2016;46(5):1593-602. doi: 10.3906/sag-1506-108.
- Hasanpour M, Ashrafi M, Erjaee H, Nazifi S. The effect of saffron aqueous extract on oxidative stress parameters and important biochemical enzymes in the testis of streptozotocininduced diabetic rats. Physiol Pharmacol. 2018;22(1):28-37.
- 39. Yaribeygi H, Mohammadi MT, Rezaee R, Sahebkar A. Crocin improves renal function by declining Nox-4, IL-18, and

p53 expression levels in an experimental model of diabetic nephropathy. J Cell Biochem. 2018;119(7):6080-93. doi: 10.1002/jcb.26806.

- Ghorbanzadeh V, Mohammadi M, Mohaddes G, Dariushnejad H, Chodari L, Mohammadi S. Protective effect of crocin and voluntary exercise against oxidative stress in the heart of high-fat diet-induced type 2 diabetic rats. Physiol Int. 2016;103(4):459-68. doi: 10.1556/2060.103.2016.4.6.
- Farshid AA, Tamaddonfard E, Moradi-Arzeloo M, Mirzakhani N. The effects of crocin, insulin and their co-administration on the heart function and pathology in streptozotocin-induced diabetic rats. Avicenna J Phytomed. 2016;6(6):658-70.
- 42. Farshid AA, Tamaddonfard E. Histopathological and behavioral evaluations of the effects of crocin, safranal and insulin on diabetic peripheral neuropathy in rats. Avicenna J Phytomed. 2015;5(5):469-78.
- 43. El-Fawal R, El Fayoumi HM, Mahmoud MF. Diosmin and crocin alleviate nephropathy in metabolic syndrome rat model: effect on oxidative stress and low-grade inflammation. Biomed Pharmacother. 2018;102:930-7. doi: 10.1016/j. biopha.2018.03.162.
- Bajerska J, Mildner-Szkudlarz S, Podgórski T, Oszmatek-Pruszyńska E. Saffron (*Crocus sativus* L.) powder as an ingredient of rye bread: an anti-diabetic evaluation. J Med Food. 2013;16(9):847-56. doi: 10.1089/jmf.2012.0168.
- 45. Bahmani F, Bathaie SZ, Aldavood SJ, Ghahghaei A. Inhibitory effect of crocin(s) on lens α-crystallin glycation and aggregation, results in the decrease of the risk of diabetic cataract. Molecules. 2016;21(2):143. doi: 10.3390/ molecules21020143.
- 46. Asri-Rezaei S, Tamaddonfard E, Ghasemsoltani-Momtaz B, Erfanparast A, Gholamalipour S. Effects of crocin and zinc chloride on blood levels of zinc and metabolic and oxidative parameters in streptozotocin-induced diabetic rats. Avicenna J Phytomed. 2015;5(5):403-12.
- Ashrafi M, Nazifi S, Namazi F, Kazemipour N, Karimi B, Goudarzi T, et al. Renal protective effect of saffron aqueous extract in streptozotocin induced diabetic rats. Int J Med Res Health Sci. 2017;6(9):151-61.
- Altinoz E, Taskin E, Oner Z, Elbe H, Arslan BA. The effect of saffron (its active constituent, crocin) on the cardiovascular complication and dyslipidemia in streptozotocin induced diabetic rats. Afr J Tradit Complement Altern Med. 2015;12(5):1-7. doi: 10.4314/ajtcam.v12i5.1.
- Altinzo E, Oner Z, Elbe H, Vardi N. Neuro-protective effects of crocin on brain and cerebellum tissues in diabetic rats. Afr J Tradit Complement Altern Med. 2014;11(6):33-9. doi: 10.4314/ajtcam.v11i6.2.
- 50. Altinoz E, Oner Z, Elbe H, Cigremis Y, Turkoz Y. Protective effects of saffron (its active constituent, crocin) on nephropathy

in streptozotocin-induced diabetic rats. Hum Exp Toxicol. 2015;34(2):127-34. doi: 10.1177/0960327114538989.

- Ahmadi M, Rajaei Z, Hadjzadeh MA, Nemati H, Hosseini M. Crocin improves spatial learning and memory deficits in the Morris water maze via attenuating cortical oxidative damage in diabetic rats. Neurosci Lett. 2017;642:1-6. doi: 10.1016/j. neulet.2017.01.049.
- 52. Abou-Hany HO, Atef H, Said E, Elkashef HA, Salem HA. Crocin mediated amelioration of oxidative burden and inflammatory cascade suppresses diabetic nephropathy progression in diabetic rats. Chem Biol Interact. 2018;284:90-100. doi: 10.1016/j.cbi.2018.02.001.
- Tamaddonfard E, Farshid AA, Asri-Rezaee S, Javadi S, Khosravi V, Rahman B, et al. Crocin improved learning and memory impairments in streptozotocin-induced diabetic rats. Iran J Basic Med Sci. 2013;16(1):91-100.
- Altinoz E, Oner Z, Elbe H, Turkoz Y, Cigremis Y. Protective effect of saffron (its active constituent, crocin) on oxidative stress and hepatic injury in streptozotocin induced diabetic rats. Gene Ther Mol Biol. 2014;16(1):160-71.
- 55. Kapucu A. Crocin ameliorates oxidative stress and suppresses renal damage in streptozotocin induced diabetic male rats. Biotech Histochem. 2021;96(2):153-60. doi: 10.1080/10520295.2020.1808702.
- Skourtis G, Krontira A, Ntaoula S, Ferlemi AV, Zeliou K, Georgakopoulos C, et al. Protective antioxidant effects of saffron extract on retinas of streptozotocin-induced diabetic rats. Rom J Ophthalmol. 2020;64(4):394-403. doi: 10.22336/ rjo.2020.61.
- 57. Yaribeygi H, Atkin SL, Barreto GE, Sahebkar A. Crocin improves oxidative stress in testicular tissues of streptozotocininduced diabetic rats. Adv Exp Med Biol. 2021;1308:273-81. doi: 10.1007/978-3-030-64872-5_19.
- Yaribeygi H, Zare V, Butler AE, Barreto GE, Sahebkar A. Antidiabetic potential of saffron and its active constituents. J Cell Physiol. 2019;234(6):8610-7. doi: 10.1002/jcp.27843.
- 59. Matough FA, Budin SB, Hamid ZA, Alwahaibi N, Mohamed J. The role of oxidative stress and antioxidants in diabetic complications. Sultan Qaboos Univ Med J. 2012;12(1):5-18. doi: 10.12816/0003082.
- Caballero-Ortega H, Pereda-Miranda R, Abdullaev FI. HPLC quantification of major active components from 11 different saffron (*Crocus sativus* L.) sources. Food Chem. 2007;100(3):1126-31. doi: 10.1016/j.foodchem.2005.11.020.
- Hosseinzadeh H, Modaghegh MH, Saffari Z. Crocus sativus L. (saffron) extract and its active constituents (crocin and safranal) on ischemia-reperfusion in rat skeletal muscle. Evid Based Complement Alternat Med. 2009;6(3):343-50. doi: 10.1093/ecam/nem125.